

1 **Limnology and diatom ecology of shallow lakes in a rapidly thawing**  
2 **discontinuous permafrost peatland**

3

4 Kristen A Coleman<sup>a\*</sup>, Grace N. Hoskin<sup>a</sup>, Laura Chasmer<sup>b</sup>, Joshua R. Thienpont<sup>a</sup>,  
5 William L. Quinton<sup>c</sup> and Jennifer B. Korosi<sup>a</sup>

6 <sup>a</sup> *Faculty of Environmental and Urban Change, York University, Toronto, Ontario, Canada, M3J 1P3*

7 <sup>b</sup> *Department of Geography and Environment, University of Lethbridge, Lethbridge, Alberta, Canada,*  
8 *TIK 3M4*

9 <sup>c</sup> *Cold Region Research Centre, Wilfrid Laurier University, Waterloo, Canada, N2L 3C5*

10

11 \*corresponding author: [kcoleman@yorku.ca](mailto:kcoleman@yorku.ca)

12

13 This is an Accepted Manuscript of an article published by Taylor & Francis in the journal Inland Waters  
14 on February 1, 2023, available online: <https://doi.org/10.1080/20442041.2022.2144699>

15 **Limnology and diatom ecology of shallow lakes in a rapidly thawing**  
16 **discontinuous permafrost peatland**

17 Lakes in discontinuous permafrost peatlands are on the front lines of climate change, sensitive to even  
18 modest increases in air temperature. The aim of this study was to provide the first limnological  
19 characterization of shallow (~1-2 m depth) lakes in the Scotty Creek basin (Northwest Territories,  
20 Canada), a field site of circumpolar significance due to the existence of long-term ecohydrological  
21 monitoring going back decades. We use this as a foundation from which to advance our process-based  
22 understanding of the potential drivers of lake ecosystem change. Our results showed that dissolved  
23 organic carbon (DOC) and lake color were not correlated, a pattern that appears to be an important driver  
24 of diatom (siliceous single-celled algae) assemblages in these lakes. Diatoms in the study lakes tended to  
25 fall into one of two assemblage clusters. One cluster, comprised of small benthic Fragilariaceae and small  
26 *Navicula* species (*sensu lato*), was found associated with higher lake color. The second cluster, comprised  
27 of *Encyonopsis* and large *Navicula* species, was found associated with high DOC, lower color, and the  
28 presence of a benthic moss mat. From this, we suggest that DOC quality is a primary control on lake  
29 ecology in this region for its role in controlling light penetration to the lake bottom. We hypothesized that  
30 the prevalence of nearshore fens and collapse scar wetlands would be important drivers of DOC, but this  
31 was not supported in the 9 study lakes for which we had available data to map shoreline features.

32

33 **Keywords:** high-resolution data, collapse scars, channel fens, climate change, lake browning, DOC

## 34 **1. Introduction**

35 Permafrost peatlands cover approximately 19% of the circumpolar permafrost region  
36 (Tarnocai et al. 2009) and contain globally significant stores of terrestrial carbon (Harris et al.  
37 2021). Climate change and an accelerated rate of permafrost thaw have raised concerns about  
38 their future role in the global carbon cycle under a warming climate (Hugelius et al. 2020).  
39 Shallow lakes are abundant features across many permafrost peatland landscapes of the  
40 circumpolar North. They play a critical role in the cycling of water and chemical constituents  
41 across the landscape, and act as both sentinels and agents of environmental change.  
42 Consequently, attempts to decipher the future role of permafrost peatlands in the global carbon  
43 balance must include an understanding of how shallow lake ecosystem structure and function are  
44 likely to be altered with climate change (Cole et al. 2007).

45 Discontinuous permafrost landscapes are highly sensitive to even modest warming of air  
46 temperatures (Kettles and Tarnocai 1999; Spence et al. 2020). Models project that 40% of the  
47 world's permafrost could be lost by 2100 CE (SOCR 2021), with sporadic discontinuous  
48 permafrost regions potentially permafrost-free by mid-century (Chasmer and Hopkinson 2017).  
49 Permafrost thaw can manifest in several different ways: it can change the thickness of the  
50 seasonally thawed (i.e. active) layer, generate a layer of talik separating the underlying  
51 permafrost from the overlying active layer, and produce thermokarst terrain (Zhang et al. 1999;  
52 Zhang et al. 2005; Frey and McClelland, 2009). While changes in active layer thickness occur  
53 slowly (press disturbance), thermokarst development from the thawing of ice-rich permafrost  
54 manifests as a rapid (or pulse) disturbance (Vonk et al. 2015). In southern permafrost peatlands,  
55 wetland thermokarst processes typically predominate, which results in the conversion of forests  
56 into wetlands (Quinton et al. 2019).

57 Wetland thermokarst alters the hydrologic connectivity of the landscape by increasing the  
58 number, duration, and depth of flow paths (Smith 2007; Karlsson 2012; Quinton et al. 2019).  
59 The hydrological connectivity of a landscape in turn exerts a strong control over transportation of  
60 sediment, organic matter, and nutrients, with shallow lakes playing a critical role in the  
61 processing of these materials along hydrological cascades. Increased areal wetland coverage and  
62 hydrological connectivity, coupled with increased availability of terrestrial organic matter  
63 liberated from thawed soils, has the potential to increase the export of terrestrial organic carbon  
64 to lakes. As terrestrial organic carbon is typically highly colored, increased export of carbon  
65 from the catchment can result in a phenomenon known as lake browning (Wauthy et al. 2018).  
66 Lake browning alters light penetration, thermal regime, and oxygen dynamics in lakes, and can  
67 fuel heterotrophic energy pathways in lake food webs (Cole et al. 2006; Porcal et al. 2009; Vonk  
68 et al. 2015; Tanentzap et al. 2017).

69 Increases in lake DOC concentrations are predicted with continued permafrost thaw  
70 (Vonk et al. 2013; Wauthy et al. 2018), yet paleolimnological reconstructions of thermokarst  
71 lakes have provided mixed results regarding trajectories of lake DOC concentrations (Vonk et al.  
72 2012; Bouchard et al. 2013; Thienpont et al. 2013; Coleman et al. 2015). This may be due, at  
73 least in part, to localized differences in catchment ecohydrological characteristics. Topography,  
74 permafrost extent, ground ice content, surficial geology, and Quaternary history have all been  
75 identified as key state factors that are likely to influence biogeochemical responses of watersheds  
76 to permafrost thaw (O'Neill et al. 2019, Vonk et al. 2019; Tank et al. 2020). Consequently,  
77 representation across broad geographic gradients is required to disentangle the trajectories and  
78 drivers of limnological change in northern lakes and make predictions at the circumpolar scale.  
79 Even within regions, responses of permafrost peatlands to warming can be variable, highlighting

80 the need for a better understanding of the processes underlying ecosystem change (Sim et al.  
81 2021). In this study, we contribute to addressing these knowledge gaps by providing the first  
82 limnological characterization of small, shallow lakes in the Scotty Creek basin of the  
83 southwestern Northwest Territories (Canada).

84 The Scotty Creek basin (61°180 N, 121°180 W) is a region of low-relief and extensive  
85 peatlands located at the northern extent of the sporadic discontinuous permafrost zone in the  
86 Taiga Plains Ecozone. Here, permafrost exists beneath treed peat plateaus, while surrounding  
87 wetlands, including channel fens and collapse scar wetlands (hereafter “collapse scars”), are  
88 predominately treeless and permafrost-free. Warming air temperatures, which has accelerated  
89 since ~1970, has resulted in the rapid loss of permafrost (Quinton et al. 2019). The region is  
90 predicted to be permafrost-free by mid-century (Chasmer and Hopkinson 2017). The Scotty  
91 Creek Research Station has supported intensive field, remote sensing, and modelling studies  
92 since the late ~1990’s, providing a long-term perspective on thaw-induced watershed changes  
93 rarely available for remote subarctic regions (Quinton et al. 2019). To date, however, very little  
94 research has been done on the many small lakes and ponds found within the Scotty Creek basin.  
95 In a recent comparison of hydrological trends in 32 circumpolar peatland basins, the Taiga Plains  
96 was unique in experiencing a widespread increase in annual basin runoff in the absence of  
97 increasing precipitation (Mack et al. 2021), a phenomenon that has been linked to accelerated  
98 wetland thermokarst (Connon et al. 2012). Clearly, the Scotty Creek basin is in a period of rapid  
99 transition, where limnological research should be prioritized.

100 The objectives of this study are to: (1) document the present-day limnology of shallow  
101 lakes in the Scotty Creek basin as a benchmark from which to examine future limnological  
102 change; and (2) examine spatial relationships between water chemistry, nearshore landscape

103 characteristics, and diatom (siliceous algae) assemblages to understand controls on limnological  
104 characteristics and make predictions about drivers of future change. To meet our objectives, we  
105 analyzed surface water chemistry and sedimentary diatom assemblages in 16 lakes in or near the  
106 Scotty Creek basin and used high-resolution aerial imagery available for 9 of the lakes to map  
107 channel fen and collapse scar area within 250 m of the lake perimeter. We also collected high-  
108 resolution logger data for depth, oxygen, light, and temperature profiles over the 2019 ice-free  
109 season in two lakes in the Scotty Creek basin that we propose as candidate sites for the  
110 establishment of a long-term lake monitoring program. Overall, this study provides a foundation  
111 for advancing our process-based understanding of the drivers of limnological change in rapidly  
112 thawing discontinuous permafrost peatlands.

113

## 114 **2. Methods**

### 115 ***2.1 Overview***

116 We selected 16 lakes to sample for water chemistry and sedimentary diatom assemblages.  
117 The 16 lakes were chosen because they are in close proximity to one another, and centered on the  
118 Scotty Creek basin, which allowed us to explore the range of limnological variability within a  
119 small geographic area (Figure 1). Of the 16 lakes, 8 were located within the Scotty Creek basin,  
120 an additional 6 were located just outside the boundaries of the Scotty Creek watershed, and 2  
121 (FS1 and FS2) were located northeast of Scotty Creek. FS1 and FS2 were slightly larger, and in a  
122 more fluvially-influenced catchment setting, compared to the other lakes.

123 For 9 of the 16 study lakes, including SC8 and the 8 lakes within the Scotty Creek basin,  
124 we were able to use previously available landscape classification data (Chasmer et al. 2014) to

125 quantify the area within 250 m of the lake shoreline that was composed of fens and collapse  
126 scars. We used the landscape classification data to explore relationships between water  
127 chemistry, diatoms, and land cover. Two of the lakes within the Scotty Creek basin (First Lake  
128 and Goose Lake) are easily accessible from the Scotty Creek Field Station, which allowed us to  
129 conduct more intensive field sampling. For these lakes, we deployed loggers and collected water  
130 and sediments from adjacent wetlands to further refine our understanding of their limnology.

## 131 **2.2 Field methods**

132 The 16 lakes were sampled from the center of the lake by helicopter in mid-July, 2018  
133 (Figure 1). That season, ice-off occurred between May 4 and 10, 2018. Conductivity and pH  
134 were measured at all sites *in situ* using a Hanna multiprobe, but oxygen measurements were not  
135 recorded due to equipment malfunction. Lake depths were measured using an acoustic depth  
136 sounder; however, depths were not used in statistical analyses because the interference of dense  
137 moss mats and macrophytes established on many lake bottoms prevented stabilized readings of  
138 the depth sounder. For many lakes, readings fluctuated by 10s of cm, which is significant for  
139 these shallow systems. For all sites, water samples were collected in two pre-sterilized 1-litre  
140 polyethylene bottles. One litre was filtered through 0.45 µm cellulose acetate filters within 24  
141 hours of collection for analysis of dissolved organic carbon (DOC) and total dissolved nitrogen  
142 (TN). Unfiltered water was poured into 250 mL glass bottles containing 0.5 mL nitric acid,  
143 including a field blank using DI water, and sent to SGS Laboratories for elemental analysis  
144 (Montreal, QC, Canada). Sediment cores were collected from the 16 lakes using a Uwitec gravity  
145 corer with an internal diameter of 8.6 cm and sectioned into 0.5 cm intervals using a modified  
146 Glew (1988) extruder. The top 0.5 cm of each core was used for the analysis of modern diatom

147 assemblages. Based on  $^{210}\text{Pb}$  dating, the top 0.5 cm represented ~2-5 years of sediment  
148 accumulation in these lakes (Coleman and Korosi 2023).

149 In June 2019, surface water and sediments were sampled from Goose Lake and First  
150 Lake at the Scotty Creek Research Station, as well as five fen sites and one collapse scar site  
151 (Figure 1D). Additionally, substrate samples (submerged vegetation, submerged peat, sediment,  
152 and grasses) were collected from shoreline environments around Goose Lake, and from a benthic  
153 moss mat in the centre of the north basin of Goose Lake at a water depth of ~1m. Water and  
154 sediment sampling followed the same methods used for the 2018 sampling of the 16 lakes,  
155 described above, except this time we were able to collect to collect dissolved oxygen data with  
156 the Hanna Multiprobe

157 Buoys equipped with HOBO Pendant wireless temperature and light data loggers, Onset  
158 HOBO U26 dissolved oxygen (DO) data loggers and Onset HOBO U20 depth loggers were  
159 deployed in both the main basin and northern basin of Goose Lake and center of First Lake  
160 (Figure 1D) on June 7<sup>th</sup> (~a month past ice-off) and removed on August 22<sup>nd</sup>, 2019. Each buoy  
161 was connected to a rope which was anchored to the sediment using 20 lb weights. An additional  
162 rope was suspended from the buoy, upon which three temperature/light loggers, and one oxygen  
163 logger were attached. The depth logger was attached to the weight to keep it stationary in the  
164 water column in relation to the sediments and was kept afloat using empty 2 L bottles.  
165 Precipitation data was collected using a Pluvio totalizing station, located at the Scotty Creek  
166 research station adjacent to Goose Lake, and was corrected for undercatch. Precipitation was  
167 recorded every 30 minutes to examine the impact of precipitation on other time-series data (e.g.  
168 light and depth).

### 169 *2.3 Laboratory analysis of water chemistry*



170 DOC and TN were analyzed on a Shimadzu-TOC5000 Series TOC-L analyzer following  
171 the combustion catalytic oxidation method and non-dispersive infrared detection. Standards were  
172 made using potassium hydrogen phthalate for instrument calibration and performance. Samples  
173 were acidified using HCl and sparged to purge DIC. True color was measured using the  
174 platinum-cobalt method on a Genesys 10S UV-Vis spectrophotometer (APHA 2011). Alkalinity  
175 was analyzed by titrating samples with 0.02N H<sub>2</sub>SO<sub>4</sub> (APHA 1998). Samples were sent to SGS  
176 Laboratories (Montreal, QC, Canada) for elemental analysis, including total phosphorus, using  
177 ICP-MS and ICP-OES. Although perchloric acid digestion followed by colorimetry is the more  
178 common for analysis of total phosphorus, good agreement has been found between this method  
179 and ICP-MS (Ivanov et al., 2012), as used here.

## 180 ***2.4 Physical Landscape Characterization***

### 181 *2.3.1 Shoreline Development and Lake Area*

182 The shoreline development (D<sub>L</sub>) ratio, an index that describes how close a lake is to a  
183 perfect circle, was calculated for all 16 study lakes. Lakes with a D<sub>L</sub> close to 1 have circular  
184 shapes, while lakes with high D<sub>L</sub> have more jagged and crenulated shapes. D<sub>L</sub> is the ratio of the  
185 perimeter of the lake to the perimeter of a circle with the same area

$$186 \quad D_L = S_L \div 2 \cdot \text{sqrt}(\pi \cdot A_o) \quad (1)$$

187 where S<sub>L</sub> is shoreline length, and A<sub>o</sub> is the area of the lake. Lakes with a higher D<sub>L</sub> have higher  
188 potential for the development of littoral communities and increase the exposure of lakes to the  
189 surrounding shorelines. Lake area and perimeter were calculated using the polygon function in  
190 Google Earth.

### 191 *2.3.2 Buffer and Overlay Landscape Analysis*

192           Nine of the 16 study lakes were used for the buffer and overlay landscape analysis.  
193   Buffering is the process of creating an output polygon layer that delineates a zone of specified  
194   width around a feature of interest. An overlay is the process of taking two or more different maps  
195   of the same area and placing them on top of one another. In this study, we applied a 250 m buffer  
196   around the perimeter of the 9 study lakes, followed by an overlay analysis to determine how  
197   much fen and collapse scar area fell within each buffer. The outputted layers included shapefiles  
198   only of the collapse scars and fens that overlapped within the 250 m buffer. The buffer and  
199   overlay analyses were done in ArcMap version 10.7.1 using a raster file of the Scotty Creek  
200   study area. Individual shapefiles were created for each land classification (channel fen, collapse  
201   scars, water, moraine/upland, and permafrost) and the water layer was separated into individual  
202   lakes.

### 203   ***2.5 Subfossil Diatom Analysis***

204           Diatom slides were prepared following Battarbee et al. (2001) using the hydrogen  
205   peroxide method and mounted using Naphrax®. Diatom valves were identified using an  
206   Amscope B690C-PL microscope and multiple reference texts (Krammer and Lange-Bertalot  
207   1997; Krammer and Lange-Bertalot 1999; Krammer and Lange-Bertalot 2000; Fallu et al. 2000)  
208   and online databases (Guiry and Guiry, 2021; Spaulding et al. 2022). Between 300 to 500 valves  
209   were counted per site, except for Fen 4 and the collapse scar site which had markedly low  
210   diversity (Hill's  $N_2 < 4$ ), where a minimum of 175 valves were counted. Diatom species were  
211   grouped based on similar ecologies (Appendix A) and were displayed as relative abundances.  
212   Chrysophyte cysts, which can form in response to changing environmental conditions, and  
213   protozoan plates, which can provide information on moisture levels, were also present on diatom

214 slides and enumerated and displayed on figures as percentages relative to the sum of all diatom  
215 valves.

## 216 *2.6 Data Analysis*

217 Data retrieved from data loggers was plotted using the ggplot package (Wickham 2019)  
218 in R (R Core Team 2020). Light absorption per cm was calculated by the equation

$$219 \qquad [100 \times (I_0 - I_z)] / I_0 \qquad (2)$$

220 where  $I_0$  is the irradiance recorded in the shallowest sensor, and  $I_z$  is the irradiance recorded in  
221 the deepest sensor divided by the number of cm between the two sensors. Anomalous negative  
222 values caused by higher irradiance values recorded in the deeper sensor than the shallower sensor  
223 were removed, and likely are a result of shading of the shallower sensor by the buoy. A LOESS  
224 smoothed line was included in the figures using the `stat_smooth` function in ggplot in R.

225 Water chemistry variables that had concentrations below detection were not included in  
226 any statistical analyses (mercury, silver, arsenic, beryllium, boron, bismuth, molybdenum, nickel,  
227 antimony, selenium, tin, thallium, zirconium, and zinc). Variables were also removed if there  
228 was evidence of contamination based on anomalously high field blank concentrations  
229 (aluminum, cadmium, cobalt, chromium, copper, iron, lead, titanium, uranium, and vanadium).  
230 Spearman's correlation analyses, used here due to the non-normal distribution of most variables,  
231 were calculated on remaining water chemistry variables to determine if there were statistically  
232 significant relationships between water chemistry variables and lake area (Area) or shoreline  
233 development ( $D_L$ ). Water chemistry variables included in the Spearman's correlation analyses  
234 included pH, dissolved organic carbon (DOC), total nitrogen, filtered (TN), true color (color),

235 phosphorus (TP), conductivity, alkalinity, calcium (Ca), potassium (K), lithium (Li), magnesium  
236 (Mg), manganese (Mn), sodium (Na), silicon (Si), and strontium (Sr).

237         Principal components analysis (PCA) was used to visualize limnological variation among  
238 lakes. Variables were standardized using the “scale” function in the vegan package (Oksanen et  
239 al. 2019) in R. A PCA was generated for all 16 lakes to examine variation between lakes with  
240 respect to Area, D<sub>L</sub>, pH, DOC, TN, color, conductivity, Li, and Mn. Area and D<sub>L</sub> were plotted  
241 passively. Highly correlated variables were removed from the PCA. For example, conductivity  
242 was correlated with alkalinity, calcium (Ca), potassium (K), magnesium (Mg), and strontium  
243 (Sr). The remaining variables were selected due to their importance in structuring biological  
244 communities (pH, DOC, TN, TP, color, and conductivity), usefulness as a lithogenic tracer (Li),  
245 or sensitivity to redox conditions (Mn). Li, TP, and Area were normalized using a log  
246 transformation, and D<sub>L</sub> and conductivity were normalized using a loglog transformation. The  
247 remaining variables were already normally distributed, and no transformation was applied. A  
248 PCA was also generated for the subset of 9 lakes used in landscape analyses, to visually examine  
249 relationships between the same water chemistry variables as above and collapse scar or fen  
250 coverage within 250 m of the lake perimeter. Collapse scar and fen coverage were plotted  
251 passively, and Mn was normalized using a log transformation.

252         Direct ordination [Redundancy analysis (RDA)] was used to evaluate relationships  
253 between diatom community composition and environmental variables. Lake area and  
254 conductivity were log transformed to normalize distribution, and species data were Hellinger  
255 transformed to standardize species assemblage data from absolute to relative values. A variance  
256 inflation factor (VIF) test was run to assess multicollinearity. Variables with high VIFs were  
257 sequentially removed from analysis until all environmental variables had VIFs below 10.

258 Variables retained in the analysis were pH, color, DOC, area, and conductivity. Significance of  
259 the RDA was determined using an ANOVA-like permutations test. Correlation analyses were  
260 conducted using the Hmisc package (Harrell 2021), and PCAs, VIFs, RDAs and permutation  
261 tests were conducted using the Vegan package (Oksanen et al. 2019) in R.

262

### 263 **3. Results**

#### 264 *3.1 High-Resolution Assessment of Goose Lake and First Lake*

265 Depth measurements obtained from depth loggers deployed in First Lake and the two  
266 main basins of Goose Lake (Figure 1D) from June 7<sup>th</sup> to August 22<sup>nd</sup> showed good agreement  
267 between all three loggers and suggested water levels of these shallow (~1 m) lakes fluctuated by  
268 ~30-40 cm throughout this period (Figures 2A, 2B, and 2C). Water levels fluctuated through  
269 June and July and began to stabilize towards the end of the recorded period. Lowest water levels  
270 occurred in early to mid-June and mid-July. Bottom dissolved oxygen (DO) measurements  
271 ranged from 5 to 12.5 mg/L in Goose Lake, in both the north basin (Figure 2A), and the main  
272 basin (Figure 2B). In First Lake, DO measurements appeared to have dropped to 0 mg/L for  
273 periods of time, however this was likely a result of the logger dropping into the sediments, as the  
274 logger recorded these anoxic conditions during periods of lower water levels (Figure 2C).

275 The consistency between temperature readings at different depths indicated that the entire  
276 water column in both lakes was mixed throughout the recorded period (Figure 2). Temperature  
277 measurements were consistent between lakes and fluctuated between 10 and 25°C throughout the  
278 recorded period. All lakes recorded warmer periods in mid-June and late July/early August. The  
279 calculated % absorption of light per cm was similar between Goose Lake and First Lake, and

280 although light penetration fluctuated through time, no clear seasonal trends were apparent  
281 (Figure 2); however, in Goose Lake, periods of higher light absorption (e.g. in late July)  
282 coincided with warmer temperatures (Figures 2A and 2B). Rainfall events occurred sporadically  
283 throughout the season, with the largest rainfall event occurring on July 18<sup>th</sup> when 12.4 mm of  
284 rain fell over an hour. Lake depth increased by ~20 cm in Goose Lake and First Lake after this  
285 rainfall event, and maximum depth occurred on July 23<sup>rd</sup> (Figure 2).

286

## 287 ***3.2 Characterization of lakes in or near the Scotty Creek Basin***

### 288 *3.2.1 Morphology and water chemistry of the 16 lakes*

289 In 2018, lake depths of the 16 surveyed lakes ranged from ~0.9 to 2.1 m; however, dense  
290 macrophyte stands or benthic moss mats likely interfered with sonar measurements, as  
291 measurement readings in some lakes did not stabilize and fluctuated by tens of centimetres. Lake  
292 area ranged from 11 to 244 ha, except for a notably larger lake (FS1) that had an area of 713 ha  
293 (Table 1). Lakes were circumneutral to slightly alkaline (pH = 7.85 – 8.75). Alkalinity ranged  
294 from 28-124 mg/L CaCO<sub>3</sub>, conductivity ranged from 55.5 to 244.2 µS/cm, DOC ranged from  
295 12.96 to 23.74 mg/L, and water color ranged from 18-463 TCU (Table 1). TN (filtered) ranged  
296 from 0.54 to 1.54 mg/L, and TP (unfiltered) ranged from 8 to 38 µg/L.

### 297 *3.2.2 Near-shore landscape characterization of 9 selected thermokarst lakes*

298 Landscape coverage of fen environments varied between 0 and 34% within 250 m of the  
299 perimeters of the 9 lakes included in the landscape characterization analyses. Collapse scars were  
300 more extensive, covering between 13.4 and 48.9% of the landscape within 250 m of the lake  
301 perimeters (Figure 3). Lakes were generally well connected through channel fen networks,

302 except for SC5 and SC8 that were more isolated. SC8 had no fen environment within 250 m of  
303 the lake, indicating it may be hydrologically isolated, and while SC5 had a fen-like environment  
304 containing grasses and sedges along the western edge of its shoreline, this fen was not  
305 hydrologically connected to a channel fen network (Figure 3).

### 306 *3.2.3 Water chemistry of wetland environments near Goose and First Lake*

307 The five fen environments had lower pH ranges than sampled lakes, being circumneutral  
308 to slightly acidic (5.97-7.40), and had higher DOC concentrations than lakes ranging from 23.08  
309 to 33.7 mg/L. In comparison to this study, Gordon et al. (2016) previously reported lower pH  
310 values (5.15-5.46), and higher DOC values (~42-49 mg/L) for fens in the Scotty Creek basin.  
311 Water color ranged from 54 to 459 TCU, alkalinity ranged from 26 to 50 mg/L CaCO<sub>3</sub>, TN  
312 ranged from 0.52 to 0.91 mg/L, and conductivity ranged from 25 to 82 μS/cm. DO ranged from  
313 2.06 to 9.27 mg/L (or 17.6 to 89.7% saturation), and was slightly lower than what had been  
314 recorded by data loggers in Goose Lake and First Lake. The one collapse scar sampled had high  
315 DOC (46.8 mg/L), which was in range of values reported by Gordon et al. (2016), and also had  
316 high color (522 TCU), low pH (5.33), low alkalinity (4 mg/L CaCO<sub>3</sub>) and low conductivity (25  
317 μS/cm). TN (0.59 mg/L) and DO (6.35 mg/L; 58.5% saturation) was within the range of water  
318 chemistry variables measured in the 5 fen environments.

### 319 *3.2.4 Analysis of relationships between water chemistry and landscape features*

320 Principal components analysis (PCA) axis 1 explained 50% of the variation in water  
321 chemistry in the 16-lake dataset, largely driven by conductivity, lithium (Li), manganese (Mn),  
322 total nitrogen (TN), and color, while PCA axis 2 explained 23% of the variation in the data set  
323 and was largely driven by DOC and TP (Figure 4A). Generally, lakes were well dispersed

324 throughout the ordination space; however, SC3, SC8, SC1, SC2, Next Lake, SC4, SC9, SC11  
325 were well dispersed along PCA axis 2, but not axis 1, indicating these lakes differed in water  
326 chemistry variables associated with axis 2 (DOC and TP). Some lakes appeared to have similar  
327 water chemistry (e.g. SC2 and Next Lake; Figure 4A), despite being relatively far apart in  
328 physical space (Figure 1). PCA ordination of the 9-lake dataset (Figure 4B) illustrated the  
329 relationship between the same water chemistry variables examined in Figure 4A and proportion  
330 of collapse scar and fen environments within 250 m of lakes. Here, PCA axis 1 explained 46% of  
331 the variation in the dataset and was driven by conductivity, Mn, TN, Li, and TP, while PCA axis  
332 2 explained 21% of the dataset and was driven by pH, color, and DOC (Figure 4B).

333 Correlation analyses suggested that lake area in this region was strongly ( $r>0.6$ ) and  
334 positively correlated to conductivity, ( $r=0.79$ ,  $p<0.01$ ), alkalinity ( $r=0.85$ ,  $p<0.01$ ), major ions  
335 such as Ca ( $r=0.74$ ,  $p<0.01$ ), Mg ( $r=0.77$ ,  $p<0.01$ ), and K ( $r=0.78$ ,  $p<0.01$ ) and moderately  
336 ( $0.3>r<0.6$ ) and positively correlated with TN ( $r=0.54$ ,  $p<0.05$ ), Mn ( $r=0.53$ ,  $p<0.05$ ), and Sr  
337 ( $r=0.57$ ,  $p<0.05$ )(Table 6). Lake area was also found to be strongly and negatively correlated to  
338 color ( $r = -0.85$ ,  $p<0.01$ ) (Table 2). Shoreline development index was moderately and positively  
339 correlated with conductivity ( $r=0.50$ ,  $p<0.05$ ) and Mg ( $r=0.48$ ,  $p<0.05$ ) (Table 2). DOC was  
340 moderately and positively correlated to both TN ( $r=0.52$ ,  $p<0.05$ ) and TP ( $r=0.58$ ,  $p<0.05$ ) (Table  
341 2). TN was not significantly correlated to TP ( $r=0.48$ ,  $p>0.05$ ) (Table 2). Color was moderately,  
342 and negatively correlated to pH ( $r=-0.54$ ,  $p<0.05$ ), and strongly and negatively related to TN  
343 ( $r=0.69$ ,  $p<0.01$ ), while TN was moderately and positively related to lithium ( $r=0.50$ ,  $p<0.05$ )  
344 (Table 2). DOC and color were not correlated in these lakes ( $r=0.16$ ,  $p=0.55$ ) (Table 2).

### 345 ***3.3 Diatom communities***

#### 346 *3.3.1 Lakes*



347           With a few exceptions, two common diatom assemblages were identified in the small  
348 lakes within the Scotty Creek basin and surrounding areas. The first assemblage, identified in  
349 SC9, SC3, SC2, SC1, SC5, and SC11, was dominated by small benthic Fragilariaceae (mainly  
350 *Pseudostaurosira brevistriata*, *P. parasitica*, *Staurosira construens*, and *S. venter*) and small  
351 *Navicula* species *sensu lato*, including *Sellaphora nigri*, and *Sellaphora seminulum* (Figure 5).  
352 The second assemblage, identified in SC4, SC6, SC12, Next Lake, FS2, Goose Lake, and SC10  
353 was dominated by *Encyonopsis* species (*E. cesatii* and *E. descripta*), and large *Navicula* species  
354 (*N. cryptocephala*, *N. cryptotenella*, and *N. radiosa*). Many of these lakes (SC12, Goose Lake,  
355 SC10, Next Lake, and FS2) also had high abundances of *Brachysira* species (*B. styriaca*, *B.*  
356 *vitrea*, and *B. zellensis*) (Figure 5). There were a few exceptions to these common groupings:  
357 The assemblage in FS1 was dominated by *Achnantheidium minutissimum*; the assemblage in SC8  
358 was dominated by *Achnantheidium minutissimum*, large *Navicula* species, *Nitzschia* species, and  
359 *Brachysira* species; and the assemblage in First Lake was dominated by *Achnantheidium*  
360 *minutissimum*, large *Navicula* species, *Nitzschia* species (mainly *N. fonticola* and *N. palea*), and  
361 centric planktonic species (*Lindavia michiganiana*, and *Discostella pseudostelligera*) (Figure 5).  
362 The abundance of chrysophyte cysts relative to diatom valves was variable, ranging from 0 to  
363 ~20%. Higher abundances were found in SC3 (~20%) and SC11 (~19%) and absent in FS1 and  
364 SC9. Protozoan plates were present in low abundance in most lakes (~0.2-4.6%) and absent in  
365 SC9 and SC10 (Figure 5).

366           Redundancy analysis (RDA) of diatom assemblages and pH, color, dissolved organic  
367 carbon, lake area, and conductivity was statistically significant ( $F=1.8422$ ,  $p<0.05$ ). The selected  
368 variables collectively explained 47.9% of the variance in the diatom assemblages, 40% of which  
369 was explained in the first two RDA axes (Figure 6). Some species preferences were identified in

370 the RDA. For example, *Brachysira vitrea* was found to be associated with lakes with greater  
371 surface area. *Navicula cryptocephala*, *N. radiosa*, *Encyonopsis cesatii*, *E. descripta*, *Kobayasiella*  
372 *jaagi*, *Cymbopleura incerta*, and *Brachysira styriaca* were all found to be associated with lakes  
373 with higher DOC, while *Staurosira construens*, *Staurosira venter*, and *Pseudostaurosira*  
374 *brevistriata* were associated with high color. Lakes SC1, SC2, SC3, SC5, and SC9 had a similar  
375 diatom assemblage, while Next Lake, Goose Lake, and lakes SC4, SC6, SC10 and SC12 had  
376 similar assemblages. FS2, SC8, and First Lake also had a similar assemblage (Figure 6).

### 377 3.3.2 Wetland diatom assemblages

378 Diatom assemblages identified in the wetland samples were variable but exhibit notable  
379 differences from lake diatom assemblages. The most abundant diatoms found in fen  
380 environments were *Eunotia* species (mainly *E. bilunaris* and *E. subarcuatooides*), *Gomphonema*  
381 species (mainly *G. parvulum*), and *Tabellaria* spp. *Rossithidium pusillum*, *Achnantheidium*  
382 *minutissimum* (Figure 7), *Nitzschia* species (mainly *N. fonticola* and *N. palea*), large *Navicula*  
383 species (mainly *N. cryptocephala*, *N. cryptotenella*, and *N. radiosa*), *Kobayasiella subtilissima*,  
384 and planktonic *Fragilaria* (mainly *F. tenera*) species were also present (Figure 7). The relative  
385 abundances of chrysophyte cysts (7-36%) and protozoan plates were variable (2-37%), but  
386 generally higher in fens than lakes (Figure 7). The diatom assemblage in the collapse scar sample  
387 was dominated by *Kobayasiella subtilissima* (95%). The highest relative abundances of both  
388 chrysophyte cysts and protozoan plates were found in the collapse scar sample (both at 42%  
389 relative abundance) (Figure 7)

### 390 3.3.3 Diatom assemblages identified in lake substrate samples

391 In the lake substrate samples, *Tabellaria* spp and large *Navicula* species (mainly *N.*  
392 *cryptocephala*, *N. cryptotenella*, and *N. radiosa*) were the most abundant diatom species  
393 identified. *Tabellaria* spp was present in high abundances in grass (~48%) and submerged  
394 vegetation (~71%) samples and was also present in the submerged peat sample (~11%) (Table  
395 3). Large *Navicula* species were found in high abundance in the sediment (~38%), submerged  
396 peat (~30%) and benthic moss mat (~39%) samples (Table 3). In addition to the large *Navicula*  
397 species, the diatom assemblage identified in the sediment sample also consisted of planktonic  
398 *Fragilaria* species (mostly *F. tenera*), *Encyonema* species (mostly *E. silesiacum*) and *Brachysira*  
399 species (*B. styriaca* and *B. vitrea*) (Table 3). The submerged peat sample also contained  
400 planktonic *Fragilaria* species, and *Encyonopsis* species (*E. descripta* and *E. cesatii*). In addition  
401 to *Tabellaia* spp. the grass sample also consisted of *Gomphonema* species (mostly *G. parvulum*)  
402 and *Achnantheidium minutissimum*, while the benthic moss mat sample consisted of large  
403 *Navicula* species, *Encyonopsis* species and *Brachysira* species (*B. styriaca* and *B. vitrea*) (Table  
404 3). Chyrosphyte cyst abundances were generally low in the substrate samples (2 to 6%), except  
405 for submerged peat, which had higher abundances (26%). Protozoan plates were almost entirely  
406 absent in these samples (~0 to 0.25%) (Table 3).

407

#### 408 **4. Discussion**

409 The peatland-dominated Scotty Creek basin is in a period of rapid transition due to  
410 climate warming and rapid loss of permafrost. The aim of this study was to characterize the  
411 present-day limnology and ecology of shallow lakes in or near Scotty Creek, and underlying  
412 drivers, as a benchmark from which to study future change. Spatial patterns in water chemistry,  
413 lake physical characteristics and diatom assemblages were explored in 16 lakes. For 9 of the 16

414 lakes where landscape classification data was available, the extent of fen and collapse scar  
415 coverage was explored as a driver of water chemistry. Lastly, loggers were deployed in First  
416 Lake and Goose Lake to gain a better understanding of seasonal dynamics in water depth,  
417 temperature, and oxygen.

#### 418 *4.1 Seasonal dynamics in Goose Lake and First Lake*

419 Goose Lake and First Lake did not exhibit thermal stratification over the 76-day period  
420 the loggers were deployed. Paleolimnological records indicate that thermal stability has been  
421 increasing in lakes across the circumpolar Arctic in response to climate change (Rühland et al.  
422 2015). Although stratification of shallow lakes is rare, it has been observed in shallow lakes with  
423 high DOC concentrations, as DOC can absorb more solar radiation and increase surface water  
424 temperatures (Bouchard et al. 2011; Deshpande et al. 2017). Shallow boreal lakes may also be  
425 more prone to periodic thermal stratification where trees provide shelter from wind mixing  
426 (Klaus et al. 2021).

427 Oxygen availability at the sediment-water interface is a fundamental control on lake  
428 biogeochemical cycling. For example, methanogenesis, the production of methane as a  
429 respiratory by-product, occurs in anoxic conditions, and may be fueled by the addition of  
430 terrestrial sources of organic carbon (Grasset et al. 2018). Increased stratification, in combination  
431 with increased bacterial consumption of high terrestrial carbon inputs, have resulted in prolonged  
432 periods of anoxia in some subarctic lakes in Northern Quebec (Deshpande et al. 2017). Small  
433 subarctic lakes in Norway similarly experienced decreasing oxygen concentrations with  
434 increasing DOC inputs (Couture et al. 2015). These changes could have important consequences  
435 for the carbon cycle as increased export of terrestrial carbon and longer periods of stratification  
436 may result in shallow subarctic lakes becoming important emitters of methane (Grasset et al.

437 2018). In contrast, high DOC subarctic lakes in Siberia had oxygen concentrations at equilibrium  
438 with the atmosphere (Shirokova et al. 2013). Oxygen levels near the sediment-water interface in  
439 Goose Lake and First Lake ranged between 5 and 12.5 mg/L and would be considered low for  
440 the protection of aquatic life (for reference, 9.5 mg/L is considered the lowest acceptable  
441 dissolved oxygen concentration for early life stages; CCME 1991). As both Goose Lake and  
442 First Lake have high DOC concentrations (23.74 and 21.93 mg/L, respectively) high rates of  
443 microbial respiration may be responsible for lower oxygen levels, as there is no evidence of  
444 stratification.

445 Peatlands cover approximately 19% of the circumpolar permafrost region (Tarnocai et al.  
446 2009), yet high resolution limnological data of these sites is scarce and is a notable gap in global  
447 lake monitoring networks such as GLEON (Global Lakes Ecological Observation Network). As  
448 permafrost wetland complexes near the southern extent of permafrost are particularly vulnerable  
449 to rising air temperatures, the Scotty Creek basin offers an ideal location for long-term  
450 limnological research. Lake monitoring data collected here can be combined with ongoing field,  
451 remote sensing, and modelling research at the Scotty Creek Research Station, to provide a more  
452 holistic understanding of changes in terrestrial-aquatic linkages with permafrost thaw.

#### 453 *4.2 Spatial patterns in lake water chemistry*

454 We expected the relative proportion of collapse scars and channel fens within 250 m of  
455 the lake shoreline would be an important driver of water chemistry in the Scotty Creek basin,  
456 because of the differential roles these landscape features play in the biogeochemical processing  
457 and transport of materials from land to lake. In this landscape, channel fens act as water  
458 conveyors, while collapse scars act predominately as water storage (Quinton et al. 2019).  
459 Collapse scars may be important sources of chromophoric DOC, as they were identified in this

460 study as having high DOC and high color (522 TCU). Our spatial results, however, showed  
461 limited evidence for the role of channel fens and collapse scars in driving lake water chemistry.  
462 For example, SC5 and SC8 were hydrologically isolated from the channel fen network, and both  
463 had high color, which may suggest that channel fens have a role in flushing terrestrial DOC from  
464 lakes; however, Next Lake also had relatively high color despite being well-connected  
465 hydrologically.

466 We postulate that temporal dynamics in a rapidly changing landscape could explain why  
467 we do not see strong spatial evidence for the importance of fen and collapse scar extent in  
468 influencing lake water chemistry. For example, an increase in fen development could initially  
469 increase color in lakes due to a phenomenon known as “bog capture” (Connon et al. 2014),  
470 where previously isolated collapse scar wetlands become connected to the expanding fen  
471 network as permafrost plateaus continue to degrade. The highly colored water is then conveyed  
472 to nearby lakes through the fen network. With time, colored DOC may ultimately be flushed  
473 from Next Lake and other lakes recently influenced by the capture of collapse scars.  
474 Paleolimnological approaches that reconstruct DOC trajectories in lakes in combination with a  
475 time series of remotely sensed images could be used in future studies to test this.

#### 476 ***4.3 Diatom ecology in permafrost wetlands and lakes of the Scotty Creek basin***

477 Two different diatom assemblages were observed in 13 of the 16 lakes sampled. The first  
478 assemblage, identified in 6 lakes, was comprised of small benthic Fragilariaceae and small  
479 *Navicula* species (*sensu lato*), which were found in this study to be associated with higher color.  
480 A similar assemblage has been identified in a lake with high color located southeast of the Scotty  
481 Creek basin near Tathlina Lake (Coleman et al. 2015; Korosi et al. 2015). Small benthic  
482 Fragilariaceae have been used as indicators of harsher, low light conditions and tend to decrease

483 in abundance with warming (Rühland et al. 2008; Bouchard et al. 2013). Although primarily  
484 benthic, they have also been identified as tychoplanktonic (e.g. Velez et al. 2021), and can be  
485 displaced into the water column with mixing; However, their absence in Goose Lake and First  
486 Lake, which appeared to be well mixed at least during the ice-free season of 2019, suggests that  
487 their presence is not related primarily to column mixing.

488         The second assemblage, recorded in 7 lakes, was comprised of *Encyonopsis* species and  
489 large *Navicula* species. These species were found to be associated with high DOC and lower pH  
490 (though note the pH gradient is narrow, ranging from 7.85 - 8.75). The species assemblages in  
491 these lakes were similar to the assemblage found in the benthic moss mat sampled from Goose  
492 Lake. While large *Navicula* species have been associated with higher DOC (Rühland et al. 2003,  
493 Coleman et al. 2015), *Encyonopsis* species are epiphytic and often associated with mosses (Bahls  
494 2013). Therefore, a main driver of diatom assemblages in these lakes may be the presence of a  
495 benthic moss mat, which is dependent on light being able to penetrate to the benthic zone of  
496 these lakes. Colored DOC could have limited the growth of moss and associated epiphytes in  
497 these lakes. DOC has previously been shown to negatively impact benthic primary production in  
498 boreal lakes in Sweden and Alaska through light limitation (Seekell et al. 2015), though the  
499 study did not distinguish between DOC and lake color and aquatic plants are known to produce  
500 autochthonous, non-chromophoric DOC (Pagano et al. 2014). We infer autochthonous DOC  
501 sources from extensive moss and macrophyte growth to be the reason why our diatom epiphyte-  
502 dominated lakes exhibited high DOC but were generally lower in color. A better understanding  
503 of the interplay between lake depth, DOC, light, and benthic primary production is needed to  
504 predict the consequences of permafrost thaw on shallow lakes in boreal permafrost peatlands.

505           The species assemblages that were present in high abundances in wetlands were different  
506 from assemblages that were identified in lakes. Three of the most common species or groups of  
507 species, were *Eunotia* species, *Gomphonema* species, and *Tabellaria* spp. *Eunotia* species in  
508 particular are commonly identified in paleoecological studies using peat cores, and have lower  
509 pH optima (e.g. Hargan et al. 2015). *Tabellaria* spp have been found in a wide variety of  
510 freshwater habitats, including lakes and peatlands, and have broad morphological variations in  
511 terms of size, but shorter species/varieties as found here have often been observed in peatlands  
512 and attached to vegetation (DeColibus, 2013). This is consistent with the assemblages identified  
513 in the substrate samples, as *Tabellaria* spp. was found in high abundances in the grasses and  
514 submerged vegetation. The diatom assemblage identified in the collapse scar site was found to be  
515 different from both lakes and fens and was dominated by *Kobayasiella subtilissima* (95%).  
516 Hargan et al. (2015) also found that *K. subtilissima* had higher prevalence in collapse scar  
517 habitats. Collapse scars also have the highest abundance of cysts and protozoan plates compared  
518 to both fens and lakes.

519           The two different assemblages of diatoms that were identified in these shallow subarctic  
520 lakes are likely driven by relative differences in benthic to pelagic production, which is impacted  
521 by the ability of light to penetrate into the water column. As increasing colored DOC  
522 concentrations can reduce light penetration, paleolimnological studies using diatoms as  
523 paleoindicators may be useful for examining the trajectory of lake browning with continued  
524 permafrost thaw. The low abundances of common wetland species also identified in lakes  
525 suggests that we are not seeing a strong biological signal of these environments in the lakes. We  
526 can therefore be confident that the diatom assemblages recorded in lakes are indicative of that  
527 lake's conditions and have not been transported from other locations. It is, however, important to



528 note that changing lake depth and duration of the ice-free season may also influence the relative  
529 importance of benthic to pelagic production in lakes and decreases in lake depth are expected  
530 with drying of wetlands with on-going thaw and loss of permafrost (Chasmer and Hopkinson  
531 2016; Carpino et al. 2021). This highlights the importance of continued high-resolution  
532 monitoring in addition to paleolimnological studies to better predict how these lakes will change  
533 with future warming in this dynamic landscape.

#### 534 ***4.4 Summary of main findings and future directions***

535         The Scotty Creek basin is experiencing a rapid loss of permafrost that manifests as  
536 wetland thermokarst, where ground subsidence results in the conversion of black spruce forest  
537 into wetlands and enhances watershed hydrological connectivity (Quinton et al. 2019). Given the  
538 differing roles that forested peat plateaus, channel fens, and collapse scars play in the transport  
539 and processing of DOC, we predicted that the degree of lake connection to the channel fen  
540 network could be related to lake chromophoric DOC, but this was not supported by our data.  
541 DOC and lake color, however, do appear to be key drivers of diatom assemblages in these lakes,  
542 with diatom assemblages tending to fall into one of two clusters. One cluster, comprised of small  
543 benthic Fragilariaceae and small *Navicula* species (*sensu lato*), was found associated with higher  
544 lake color, while the second cluster was comprised of epiphytic *Encyonopsis* and large *Navicula*  
545 species and was associated with high DOC, lower color, and the presence of a benthic moss mat.  
546 Future directions should explore DOC quality (and its drivers) in these lakes, and the interplay  
547 between DOC, lake depth, and light in structuring diatom assemblages and benthic primary  
548 production. Furthermore, as Scotty Creek is already in a period of rapid transition, long-term  
549 studies are needed to understand the trajectories and mechanisms of limnological change. First  
550 Lake and Goose Lake present ideal candidate sites for the establishment of a long-term

551 limnological monitoring program, and the close association between diatoms and lake  
552 DOC/color indicate that diatoms will be useful paleoecological indicators of changes in lake  
553 carbon and primary production dynamics in future studies.

554

#### 555 Acknowledgements

556 We would like to thank Arnab Shuvo for assistance in the field, Kristine Haynes and  
557 Mason Dominico for providing precipitation data and Emily Stewart for reviewing this  
558 manuscript and providing advice. This research was supported by a Natural Sciences and  
559 Engineering Research Council of Canada (NSERC) Discovery and Northern Supplement to JBK,  
560 and an NSERC postgraduate scholarship and Weston Family Foundation scholarship to KAC.  
561 Fieldwork funding and logistical support was also provided by ArcticNet, the Polar Continental  
562 Shelf Program and the Northern Scientific Training Program. Scotty Creek is within the  
563 traditional territory of the Liidlii Kue First Nation (LKFN). Mahsi to the LKFN and the Dehcho  
564 First Nations for their support and leadership in scientific research at Scotty Creek.

565

566 The authors report that there are no competing interests to declare

567

568 Data associated with this manuscript are available through the York University Yorkspace  
569 Institutional Repository at <http://hdl.handle.net/10315/39723>

570

571 References

- 572 APHA. 1998. Standard Methods for the Examination of Water and Wastewater. 20th Edition,  
573 American Public Health Association, American Water Works Association and Water  
574 Environmental Federation, Washington DC.
- 575 APHA. 2011. Standard methods for the examination of water and wastewater, 22nd edn.  
576 American Public Health Association, Washington, DC
- 577 Bahls L. 2013. *Encyonopsis* from western North America: 31 species from Alberta, Idaho,  
578 Montana, Oregon, South Dakota, and Washington, including 17 species described as  
579 new. Northwest Diatoms, Volume 5. The Montana Diatom Collection, Helena, 46 pp.
- 580 Battarbee RW, Jones VJ, Flower RJ, Cameron NG, Bennion H, Carvalho L, Juggins S.  
581 2001. Diatoms. In: Smol J.P., Birks H.J.B. & Last W.M. (eds.), Tracking environmental  
582 change using lake sediments, vol. 3: Terrestrial, algal, and siliceous indicators, Kluwer  
583 Academic Press, Dordrecht, The Netherlands, pp. 155–202.
- 584 Bouchard F, Francus P, Pienitz R, Laurion I. 2011. Sedimentology and geochemistry of  
585 thermokarst ponds in discontinuous permafrost, subarctic Quebec, Canada. Journal of  
586 Geophysical Research: Biogeosciences, 116(G2).
- 587 Bouchard F, Pienitz R, Ortiz JD, Francus P, Laurion I. 2013. Palaeolimnological conditions  
588 inferred from fossil diatom assemblages and derivative spectral properties  
589 of sediments in thermokarst ponds of subarctic Quebec, Canada. Boreas 42: 575–595.
- 590 Carpino O, Haynes K, Connon R, Craig J, Devoie É, Quinton W. 2021. Long-term  
591 climate-influenced land cover change in discontinuous permafrost peatland  
592 complexes. Hydrol. Earth Syst. Sci. 25(6):3301-3317.
- 593 CCME, Canadian Council of Ministers of the Environment. 1999. Canadian water quality

594 guidelines for the protection of aquatic life: Dissolved oxygen (freshwater). In: Canadian  
595 environmental quality guidelines, 1999, Canadian Council of Ministers of the  
596 Environment, Winnipeg.

597 Chasmer L, Hopkinson C, 2017. Threshold loss of discontinuous permafrost and landscape  
598 evolution. *Glob. Change Biol.* 23(7): 2672-2686.

599 Chasmer L, Hopkinson C, Veness T, Quinton W, Baltzer J. 2014. A decision-tree  
600 classification for low-lying complex land cover types within the zone of discontinuous  
601 permafrost. *Remote Sens. Environ.* 143: 73-84.

602 Cole JJ, Carpenter SR, Pace ML, Van de Bogert MC, Kitchell JL, Hodgson JR. 2006.  
603 Differential support of lake food webs by three types of terrestrial organic carbon.  
604 *Ecology Letters*, 9: 558-568.

605 Cole JJ, Prairie YT, Caraco NF, McDowell WH, Tranvik LJ, Striegl RG, Duarte  
606 CM, Kortelainen P, Downing JA, Middelburg JJ, Melack J. 2007. Plumbing the global  
607 carbon cycle: integrating inland waters into the terrestrial carbon budget. *Ecosystems*, 10:  
608 172-185.

609 Coleman KA, Korosi JB. 2023. Dataset: (Paleo)limnological data for small, shallow lakes at or  
610 near the Scotty Creek Research Station in the Dehcho region (Northwest Territories,  
611 Canada). <http://hdl.handle.net/10315/39723>

612 Coleman KA, Palmer MJ, Korosi JB, Kokelj SV, Jackson K, Hargan KE, Courtney  
613 Mustaphi CJ, Thienpont JR, Kimpe LE, Blais JM, Pisaric MFJ, Smol JP. 2015. Tracking  
614 the impacts of recent warming and thaw of permafrost peatlands on aquatic ecosystems: a  
615 multi-proxy approach using remote sensing and lake sediments. *Boreal Environ Res*, 20:  
616 363-377.

617 Connon RF, Quinton WL, Craig JR, Hayashi M. 2014. Changing hydrologic connectivity due to  
618 permafrost thaw in the lower Liard River valley, NWT, Canada. *Hydrol. Process.* 28:  
619 4163–4178.

620 Couture RM, de Wit HA, Tominaga K, Kiuru P, Markelov I. 2015. Oxygen dynamics  
621 in a boreal lake responds to long-term changes in climate, ice phenology, and DOC  
622 inputs. *J. Geophys. Res. Biogeosci.* 120: 2441–2456.

623 DeColibus D. 2013. *Tabellaria flocculoda*. In *Diatoms of North America*. Retrieved July 10,  
624 2021, from [https://diatoms.org/species/tabellaria\\_flocculosa](https://diatoms.org/species/tabellaria_flocculosa)

625 Deshpande BN, Maps F, Matveev A, Vincent WF. 2017. Oxygen depletion in subarctic  
626 peatland thaw lakes. *Arct. Sci.* 1: 1-23

627 Disher BS, Connon RF, Haynes KM, Hopkinson C, Quinton WL. 2021. The  
628 hydrology of treed wetlands in thawing discontinuous permafrost regions. *Ecohydrology*,  
629 p.e2296.

630 Fallu MA, Allaire N, Pienitz R. 2002. Distribution of freshwater diatoms in 64 Labrador  
631 (Canada) lakes: species-environment relationships along latitudinal gradients  
632 and reconstruction models for water color and alkalinity. *Can. J. Fish. Aquat. Sci.* 59:  
633 329–349

634 Frey KE, McClelland JW. 2009. Impacts of permafrost degradation on arctic river  
635 biogeochemistry. *Hydrol Process.*, 23(1): 169-182.

636 Glew JR. 1988. A portable extruding device for close interval sectioning of unconsolidated  
637 core samples. *J. Paleolimnol.* 1(3): 235-239.

638 Gordon J, Quinton W, Branfireun BA, Olefeldt, D. 2016. Mercury and methylmercury  
639 biogeochemistry in a thawing permafrost wetland complex, Northwest Territories,

640 Canada. *Hydrol Process.*, 30: 3627-3638.

641 Guiry MD, Guiry GM. 2021. *AlgaeBase*. World-wide electronic publication, National  
642 University of Ireland, Galway. <https://www.algaebase.org>; searched on 06 July 2021.

643 Hargan KE., Rühland KM, Paterson AM, Finkelstein SA, Holmquist JR, MacDonald GM, Keller  
644 W, Smol JP. 2015. The influence of water-table depth and pH on the spatial distribution  
645 of diatom species in peatlands of the Boreal Shield and Hudson Plains, Canada. *Botany*,  
646 93: 57–74.

647 Harrell FE. 2021. Hmisc: Harrell Miscellaneous. R package version 4.5-0. [https://CRAN.R-](https://CRAN.R-project.org/package=Hmisc)  
648 [project.org/package=Hmisc](https://CRAN.R-project.org/package=Hmisc)

649 Harris LI, Richardson K, Bona KA, Davidson SJ, Finkelstein SA, Garneau M, McLaughlin J,  
650 Nwaishi F, Olefeldt D, Packalen M., Roulet NT. 2022. The essential carbon service  
651 provided by northern peatlands. *Front. Ecol. Environ.*, 20: 222-230.

652 Hugelius G, Loisel J, Chadburn S, Jackson RB, Jones M, MacDonald G, Marushchak M,  
653 Olefeldt D, Packalen M, Siewert MB, Treat C. 2020. Large stocks of peatland carbon and  
654 nitrogen are vulnerable to permafrost thaw. *Proc. Nat. Acad. Sci.*, 117: 20438-20446.

655 Ivanov K, Zapryanova P, Petkova M, Stefanova V, Kmetov V, Georgieva D,  
656 Angelova V. 2012. Comparison of inductively coupled plasma mass spectrometry and  
657 colorimetric determination of total and extractable phosphorus in soils. *Spectrochimica*  
658 *Acta Part B: At. Spectrosc.*, 71: 117-122.

659 Karlsson JM, Lyon SW, Destouni G. 2012. Thermokarst lake, hydrological flow and water  
660 balance indicators of permafrost change in Western Siberia. *J. Hydrol.*464: 459–466.

661 Kettles I, and Tarnocai C. 1999. Development of a model for estimating the sensitivity of

662 Canadian peatlands to climate warning. *Géographie physique et Quaternaire*, 53(3),  
663 pp.323-338.

664 Klaus M, Karlsson J, Seekell D. 2021. Tree line advance reduces mixing and oxygen  
665 concentrations in arctic-alpine lakes through wind sheltering and organic carbon supply.  
666 *Glob. Change Biol.*, 27, 4238– 4253.

667 Korosi JB, McDonald J, Coleman KA, Palmer MJ, Smol JP, Simpson MJ, Blais  
668 JM. 2015. Long-term changes in organic matter and mercury transport to lakes in the  
669 sporadic discontinuous permafrost zone related to peat subsidence. *Limnol*  
670 *Oceanogr.* 60(5): 1550-1561.

671 Krammer K, Lange-Bertalot H. 1997. *Bacillariophyceae 2. Teil: Bacillariaceae,*  
672 *Epithemiaceae, Surirellaceae.* Süßwasserflora von Mitteleuropa 2/2. Spektrum  
673 Akademischer Verlag, Berlin, Germany.

674 Krammer K, Lange-Bertalot H. 1999. *Bacillariophyceae 1. Teil: Naviculaceae.*  
675 *Süßwasserflora von Mitteleuropa 2/1.* Spektrum Akademischer Verlag, Berlin, Germany.

676 Krammer K, Lange-Bertalot H. 2000. *Bacillariophyceae 3. Teil: Centrales, Fragilariaceae,*  
677 *Eunotiaceae.* Süßwasserflora von Mitteleuropa 2/3. Spektrum Akademischer  
678 Verlag, Berlin, Germany.

679 Mack M, Connon R, Makarieva O, McLaughlin J, Nesterova N., Quinton, W. 2021.  
680 Heterogenous runoff trends in peatland-dominated basins throughout the circumpolar  
681 North. *Environ. Res. Comms.*, 3: 075006.

682 Michelutti N, Holtham AJ, Douglas MSV, Smol JP. 2003. Periphytic diatom assemblages  
683 from ultra-oligotrophic and UV transparent lakes and ponds on Victoria Island and  
684 comparisons with other diatom surveys in the Canadian arctic. *J. Phycol.* 39: 465–480.

685 O'Neill HB, Wolfe SA, Duchesne C. 2019. New ground ice maps for Canada using a  
686 paleogeographic modelling approach. *Cryosphere* 13: 753–773.

687 Oksanen J, Blanchet FG, Friendly M, Kindt R, Legendre P, McGlinn D, Minchin PR,  
688 O'hara RB, Simpson GL, Solymos P, Stevens MHH. 2019. *Vegan: community ecology*  
689 *package (version 2.5-6). The Comprehensive R Archive Network.*

690 Pagano T, Bida M, Kenny J. 2014. Trends in Levels of Allochthonous Dissolved Organic Carbon  
691 in Natural Water: A Review of Potential Mechanisms under a Changing Climate. *Water*.  
692 6: 2862–2897.

693 Pienitz R, Smol JP, MacDonald G.M. 1999. Paleolimnological reconstruction of Holocene  
694 climatic trends from two boreal treeline lakes, Northwest Territories, Canada. *Arct.*  
695 *Antarct. Alp. Res.* 31: 82-93.

696 Porcal P, Koprivnjak JF, Molot LA, Dillon PJ. 2009. Humic substances—part 7: the  
697 biogeochemistry of dissolved organic carbon and its interactions with climate change.  
698 *Environ. Sci. Pollut. Res.* 16: 714–726.

699 Quinton W, Berg A, Braverman M, Carpino O, Chasmer L, Connon R, Craig J, Devoie  
700 É, Hayashi M, Haynes K, Olefeldt D. 2019. A synthesis of three decades of hydrological  
701 research at Scotty Creek, NWT, Canada. *Hydrol. Earth Syst. Sci.* 23(4): 2015-2039.

702 Quinton WL, Hayashi M, Chasmer LE. 2011. Permafrost-thaw-induced land-cover change in the  
703 Canadian subarctic: implications for water resources. *Hydrol. Process.* 25, 152–158.

704 R Core Team. 2020. *R: A language and environment for statistical computing.* Vienna: R  
705 foundation for Statistical Computing.

706 Rühland K, Priesnitz A, Smol JP. 2003. Paleolimnological evidence from diatoms for recent  
707 environmental changes in 50 lakes across Canadian Arctic treeline. *Arct. Antarct. Alp.*



708 Res. 35(1): 110-123.

709 Rühland K, Paterson AM, Smol JP. 2015. Diatom assemblage responses to warming: reviewing  
710 the evidence. *J. Paleolimnology* 54: 1-35

711 Seekell DA, Lapierre JF, Ask J, Bergström AK, Deininger A, Rodríguez P,  
712 Karlsson J. 2015. The influence of dissolved organic carbon on primary production in  
713 northern lakes. *Limnol. Oceanogr. Lett.* 60(4): 1276-1285.

714 Shirokova LS, Pokrovsky OS, Kirpotin SN, Desmukh C, Pokrovsky BG, Audry S, Viers J. 2013.  
715 Biogeochemistry of organic carbon, CO<sub>2</sub>, CH<sub>4</sub>, and trace elements in thermokarst water  
716 bodies in discontinuous permafrost zones of Western Siberia. *Biogeochemistry*. 113:  
717 573–593.

718 Sim TG, Swindles GT, Morris PJ, Baird AJ, Cooper CL, Gallego-Sala AV, Charman DJ, Roland  
719 TP, Borken W, Mullan DJ, Aquino-Lopez MA. 2021. Divergent responses of permafrost  
720 peatlands to recent climate change. *Environ. Res. Lett.*, 16: 034001.

721 Smith LC, Pavelsky TM, MacDonald GM, Shiklomanov AI, Lammers RB. 2007. Rising  
722 minimum daily flows in northern Eurasian rivers: A growing influence of groundwater in  
723 the high-latitude hydrologic cycle. *J. Geophys. Res. Biogeosci.* 112

724 Spaulding SA, Potapova MG, Bishop IW, Lee SS, Gasperak TS, Jovanoska E, Furey PC, Edlund  
725 MB. 2021. Diatoms. org: supporting taxonomists, connecting communities. *Diatom*  
726 *Research*. 36(4): 291-304

727 Spence C, Norris M, Bickerton G, Bonsal BR, Brua R, Culp JM, Dibike Y, Gruber S,  
728 Morse PD, Peters DL, Shrestha R. 2020. The Canadian Water Resource Vulnerability  
729 Index to Permafrost Thaw (CWRVIPT). *Arct. Sci.*, 6(4), pp.437-462

730 SOCR, State of the Cryosphere Report: A needed decade of urgent action. 2021.

731 [www.iccinet.org/statecryo21](http://www.iccinet.org/statecryo21)

732 Tanentzap AJ, Kielstra BW, Wilkinson GM, Berggren M, Craig N, Del Giorgio PA, Grey J,  
733 Gunn JM, Jones SE, Karlsson J, Solomon CT, Pace ML. 2017. Terrestrial support of lake  
734 food webs: Synthesis reveals controls over cross-ecosystem resource use. *Sci. Adv.*, 3:  
735 e1601765.

736 Tank SE, Vonk JE, Walvoord MA, McClelland JW, Laurion I, Abbott BW. 2020. Landscape  
737 matters: Predicting the biogeochemical effects of permafrost thaw on aquatic networks  
738 with a state factor approach. *Permafr. Periglac. Process.* 31: 358-370.

739 Tarnocai C, Canadell CJG, Schuur EAG, Kuhry P, Mazhitova G, Zimov S. 2009. Soil organic  
740 carbon pools in the northern circumpolar permafrost region. *Global Biogeochem. Cycles.*  
741 23: GB2023,

742 Thienpont JR, Rühland KM, Pisaric MF, Kokelj SV, Kimpe LE, Blais JM, Smol JP.  
743 2013. Biological responses to permafrost thaw slumping in Canadian Arctic lakes.  
744 *Freshw. Biol.* 58: 337-353.

745 Vonk JE, Alling V, Rahm L, Mörth CM, Humborg C, Gustafsson Ö. 2012. A centennial  
746 record of fluvial organic matter input from the discontinuous permafrost catchment of  
747 Lake Torneträsk. *J. Geophys. Res.: Biogeosci.* 117 (G3).

748 Vonk JE, Mann PJ, Davydov S, Davydova A, Spencer RGM, Schade, J., Sobczak, W.V., Zimov,  
749 N., Zimov, S., Bulygina, E., Eglinton, T.I., Holmes, R.M., 2013. High biolability of  
750 ancient permafrost carbon upon thaw. *Geophys. Res. Lett.* 40: 2689–2693.

751 Vonk JE, Tank SE, Bowden WB, Laurion I, Vincent WF, Alekseychik P, Amyot M,  
752 Billet MF, Canario J, Cory RM, Deshpande BN. 2015. Reviews and syntheses: Effects of  
753 permafrost thaw on Arctic aquatic ecosystems. *Biogeosciences.* 12(23): 7129-7167.

754 Vonk JE, Tank SE, Walvoord MA. 2019. Integrating hydrology and biogeochemistry across  
755 frozen landscapes. *Nat. Comm.* 10: 1-4.

756 Wauthy M, Rautio M, Christoffersen KS, Forsström L, Laurion I, Mariash HL, Peura  
757 S, Vincent WF. 2018. Increasing dominance of terrigenous organic matter in circumpolar  
758 freshwaters due to permafrost thaw: Increasing allochthony in arctic freshwaters. *Limnol*  
759 *Oceanogr Letters* 3: 186–198.

760 Wickham et al., (2019). Welcome to the tidyverse. *Journal of Open Source Software*, 4(43),  
761 1686

762 Zhang T, Barry RG, Knowles K, Heginbottom JA, Brown J. 1999. Statistics and  
763 characteristics of permafrost and ground ice distribution in the Northern Hemisphere.  
764 *Polar Geography*, 23(2): 132–154.

765 Zhang TJ, Frauenfeld OW, Serreze MC, Etringer A, Oelke C, McCreight J, Barry RG,  
766 Gilichinsky D, Yang DQ, Ye HC, Ling F, Chudinova S. 2005. Spatial and temporal  
767 variability in active layer thickness over the Russian Arctic drainage basin. *J Geophys*  
768 *Res- Atmospheres*, 110: D16101.

769  
770

771 Table 1: Physical characteristics and modern water chemistry statistics for 16 lakes in the Scotty  
 772 Creek and surrounding basins, collected July 2018.

	<b>Mean</b>	<b>± SD</b>	<b>Min</b>	<b>Max</b>	<b>Median</b>
Depth (m)	1.3	0.4	0.9	2.1	1.2
Area (km <sup>2</sup> )	1.06	1.69	0.11	7.13	0.32
Shoreline Development Index	1.18	0.21	1.02	1.75	1.09
pH	8.23	0.27	7.85	8.75	8.20
DOC (mg/L)	18.79	3.13	12.96	23.74	18.35
True Color (TCU)	176	124	18	463	174
TP (µg/L)(unfiltered)	18.3	7.9	8.0	38.0	17.0
TN (mg/L)(filtered)	0.90	0.28	0.54	1.54	0.79
Conductivity (µS/cm)	106.9	58.5	55.5	244.2	83.1
Alkalinity (CaCO <sub>3</sub> )(mg/L)	54	29	28	124	43
Calcium (mg/L)	15.4	6.5	7.9	29.8	13.3
Potassium (mg/L)	0.61	0.65	0.19	2.49	0.34
Lithium (µg/L)	1.4	0.8	0.7	3.2	1.2
Magnesium (mg/L)	4.67	3.08	2.24	12.70	3.51
Manganese (µg/L)	17.3	8.3	4.9	35.1	15.5
Sodium (mg/L)	2.35	1.93	0.74	7.25	1.62
Silicon (mg/L)	1.52	0.98	0.27	4.33	1.21
Strontium (µg/L)	61.1	36.0	28.5	149.0	45.3

773 SD = standard deviation; Area = surface area; DOC = dissolved organic carbon; TP = total

774 phosphorus; TN = total dissolved nitrogen

775

776 Table 2: Spearman's correlation coefficients of water chemistry variables and physical lake  
 777 characteristics for 16 lakes in the Scotty Creek basin. \*p<0.05, \*\*p<0.01

778

	Area	D <sub>L</sub>	pH	DOC	TP	TN	Color	Cond	Alk	Ca	K	Li	Mg	Mn	Na	Sr
<b>Area</b>	1.00	0.45	0.21	-0.06	-0.07	0.54	-0.85**	0.79**	0.85**	0.74**	0.78**	0.29	0.77**	0.52	0.36	0.57*
<b>D<sub>L</sub></b>		1.00	-0.07	0.12	-0.19	0.22	-0.23	0.50*	0.47	0.43	0.35	0.35	0.48	0.12	0.30	0.42
<b>pH</b>			1.00	-0.16	0.10	0.45	-0.54	0.48	0.46	0.37	0.63**	0.16	0.51*	0.48	0.42	0.49
<b>DOC</b>				1.00	0.58*	0.52*	0.16	-0.01	-0.06	-0.01	-0.06	0.17	-0.04	0.22	-0.02	-0.04
<b>TP</b>					1.00	0.48	-0.06	-0.21	-0.11	-0.28	-0.05	0.20	-0.20	0.51*	0.29	-0.21
<b>TN</b>						1.00	-0.69**	0.53	0.52*	0.49	0.65**	0.50*	0.51*	0.52*	0.53*	0.58*
<b>Color</b>							1.00	-0.76**	-0.81**	-0.68**	-0.84**	-0.38	-0.74**	-0.54*	-0.55*	-0.65**
<b>Cond</b>								1.00	0.94**	0.95**	0.83**	0.32	0.99**	0.54*	0.33	0.80**
<b>Alk</b>									1.00	0.89**	0.85**	0.36	0.95**	0.57*	0.48	0.76**
<b>Ca</b>										1.00	0.74**	0.37	0.94**	0.47	0.28	0.86**
<b>K</b>											1.00	0.19	0.84**	0.43	0.44	0.75**
<b>Li</b>												1.00	0.29	0.29	0.82**	0.55*
<b>Mg</b>													1.00	0.52*	0.37	0.77**
<b>Mn</b>														1.00	0.27	0.29
<b>Na</b>															1.00	0.51*
<b>Sr</b>																1.00

779 Area = Surface area; D<sub>L</sub> = shoreline development index; DOC = dissolved organic carbon; TP =  
 780 total phosphorus; TN = total dissolved nitrogen; Color = true color; Cond = conductivity; Alk =  
 781 alkalinity; Ca = calcium; K = potassium; Li = lithium; Mg = magnesium; Mn = manganese; Na =  
 782 sodium; Sr = strontium

783

784 Table 3: Dominant diatom species identified in lake substrate samples. Percentage of  
 785 chrysophyte cysts and protozoan plates, in relation to diatom valves, also included.

	Sediment	Submerged Peat	Grasses	Submerged vegetation	Benthic Moss Mat
<b>Location</b>	Shoreline	Shoreline	Shoreline	Shoreline	~ 1 m depth, north basin
	Large <i>Navicula</i> (~38%)	Large <i>Navicula</i> (~30%)	<i>Tabellaria</i> spp (~48%)	<i>Tabellaria</i> spp (~71%)	Large <i>Navicula</i> spp (~39%)
	Planktonic <i>Fragilaria</i> (~9%)	<i>Tabellaria</i> spp (~11%)	<i>Gomphonema</i> spp (~21%)		<i>Encyonopsis</i> spp (~14%)
<b>Dominant Diatoms</b>	<i>Encyonema</i> spp (~8%)	Planktonic <i>Fragilaria</i> (~9%)	<i>Achnantheidium minutissimum</i> (~11%)		<i>Brachysira</i> spp (~11%)
	<i>Brachysira</i> (~7%)	<i>Encyonopsis</i> spp (~9%)			<i>Achnanthes</i> spp (~10%)
<b>Cysts</b>	Cysts (%): 4%	Cysts (%): 26%	Cysts (%): 2%	Cysts (%): 2%	Cysts (%): 6%
<b>Protozoan Plates</b>	Plates (%): 0.24%	Plates (%): 0%	Plates (%): 0%	Plates (%): 0%	Plates (%): 0.25%

786

787

788 **Figure Captions**

789 Figure 1. Map of study area: the Scotty Creek Basin, Northwest Territories, Canada. (A)  
790 Approximate location of site within Canada, (B) approximate location of site within Taiga Plains  
791 ecozone, (C) location of all study lakes, (D) true color image using Sentinel 2A data (accessed  
792 June 2021) of study lakes highlighted in (C) and includes wetland sample sites (white dots).  
793 Yellow markers are locations of data loggers.

794  
795 Figure 2. Lake depth, temperature, dissolved oxygen and light absorption recorded every 30  
796 minutes from June 7<sup>th</sup> to August 22<sup>nd</sup> 2019 from Goose Lake's north basin (A), Goose Lake's  
797 main basin (B) and First Lake (C). Precipitation data was recorded from Pluvio totalizing station  
798 located near Goose Lake.

799  
800 Figure 3. Land classifications of the Scotty Creek basin based on classifications established in  
801 Chasmer et al. (2014). 250 m buffer zones are added to each lake, with the bog and fen  
802 environments within the buffer zones shown in red and yellow, respectively. Inset shows an  
803 example of the 250 m buffer zone around lake SC3, in detail.

804  
805 Figure 4: Principal components analysis (PCA) of select water chemistry variables and physical  
806 characteristics of 16 shallow lakes in the Scotty Creek basin (A) and select water chemistry  
807 variables and areal proportion of collapse scar environment (Bog250m) and fen environment  
808 (Fen250m) within 250 m of lakes shorelines for 9 lakes included in landscape classification  
809 analyses (B). Dashed lines represent variables plotted passively. Area = lake area,  $D_L$  = shoreline  
810 development index.

811

812 Figure 5: Relative abundance diagram displaying the most common diatom taxa in the modern  
813 sediments retrieved from 16 shallow lakes in the Scotty creek and neighbouring basins. Species  
814 were grouped based on similar ecology. Species or ecological groups present at less than 2%  
815 relative abundance are not shown. Lakes were ordered using diatom PCA axes scores. Ratio of  
816 chrysophyte cysts to diatom valves and protozoan plates to diatom valves are also displayed.

817

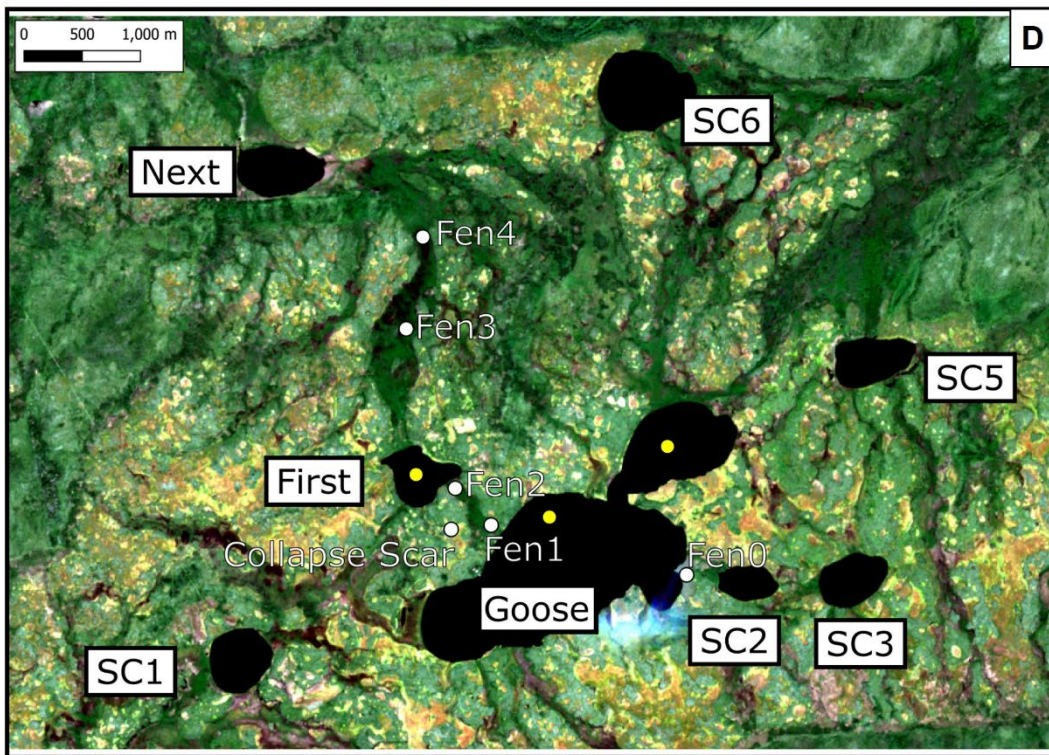
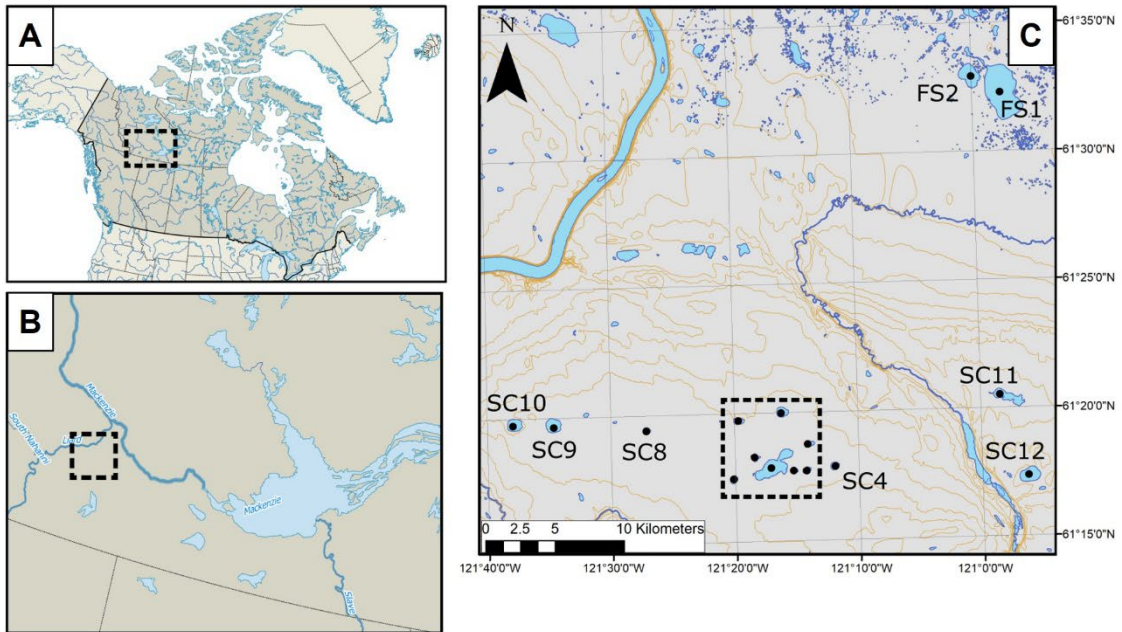
818 Figure 6: Redundancy analysis (RDA) ordination triplot illustrating relationships between diatom  
819 species and environmental variables. Environmental variables are represented by arrows. Lake  
820 sites are in black. Diatoms species codes (red) are listed in Appendix A.

821

822 Figure 7: Ground-level photographs of wetland sites and corresponding list of dominant diatom  
823 species identified in a sample retrieved from the site. Percentage of chrysophyte cysts and  
824 protozoan plates, in relation to diatom valves, are also listed.

825



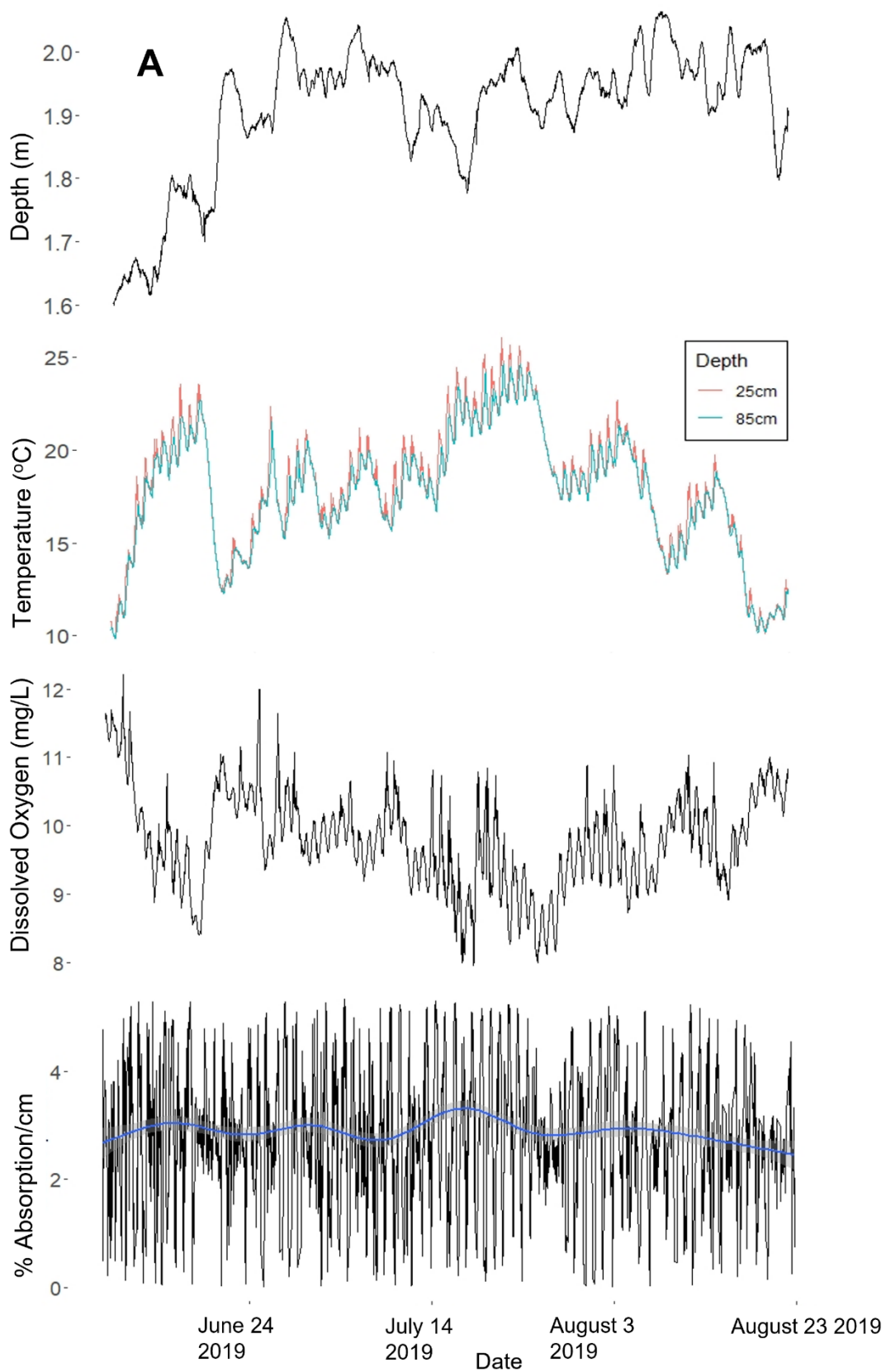


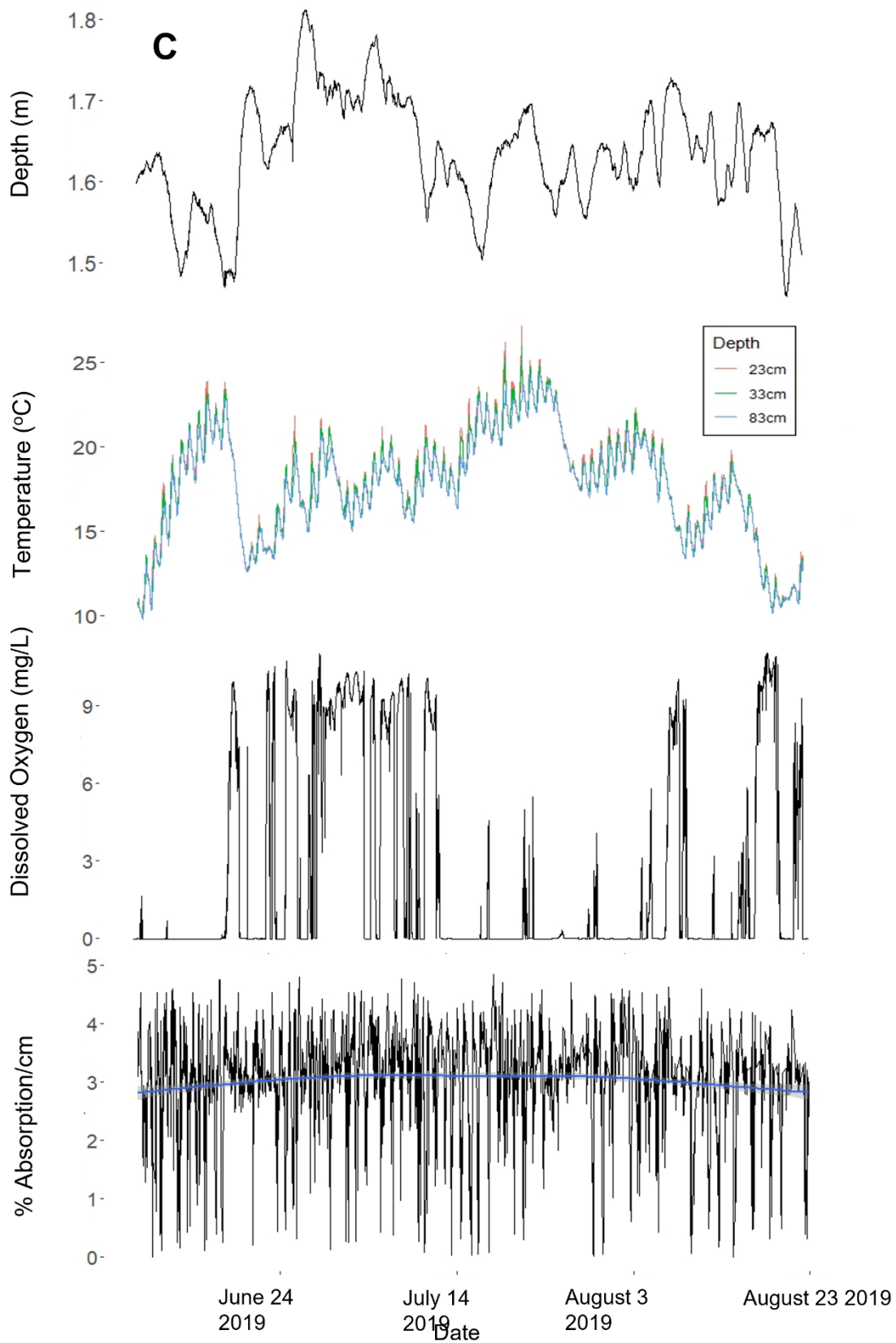
826

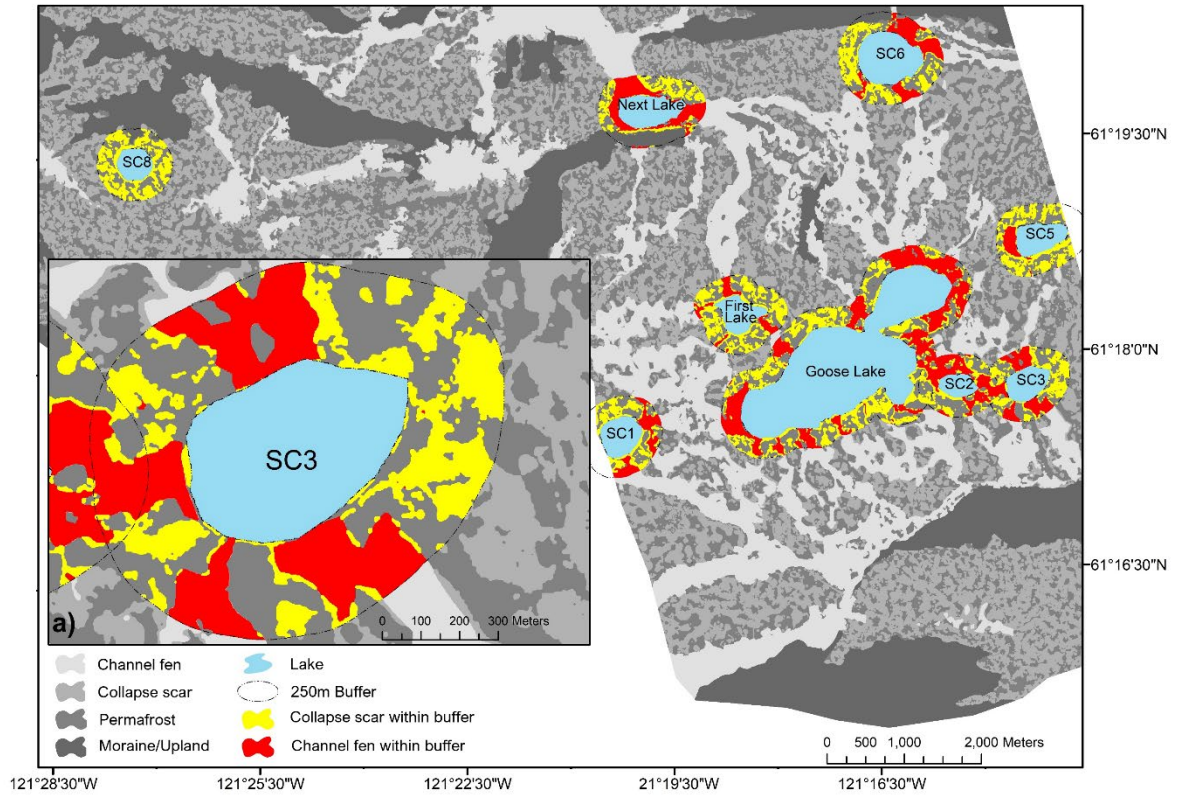
827

828

829







832

833

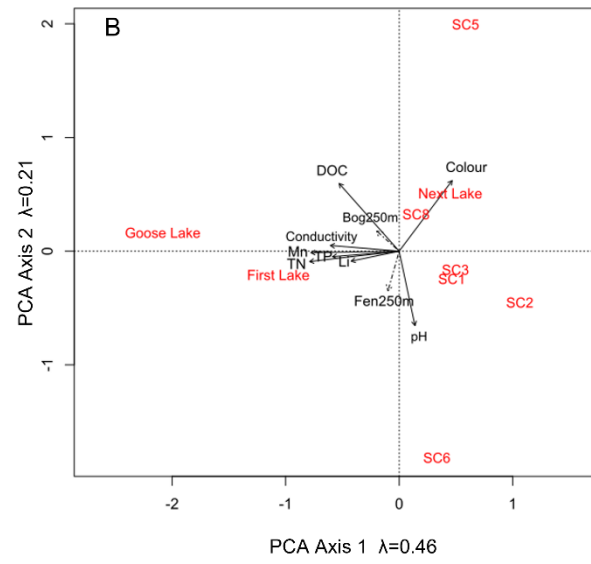
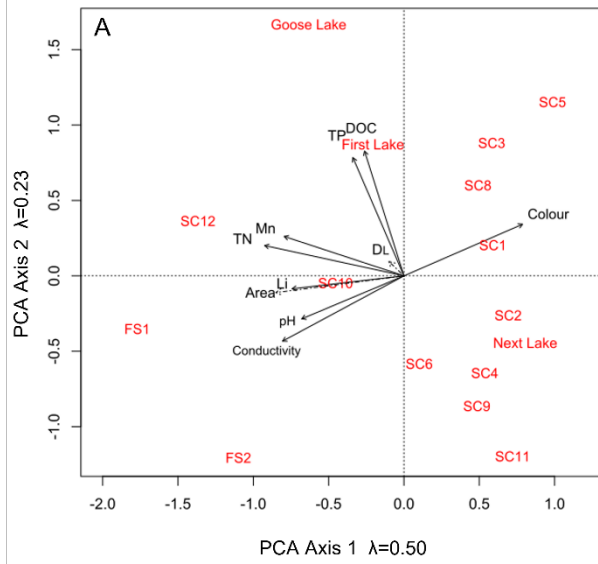
834

835

836

837

838



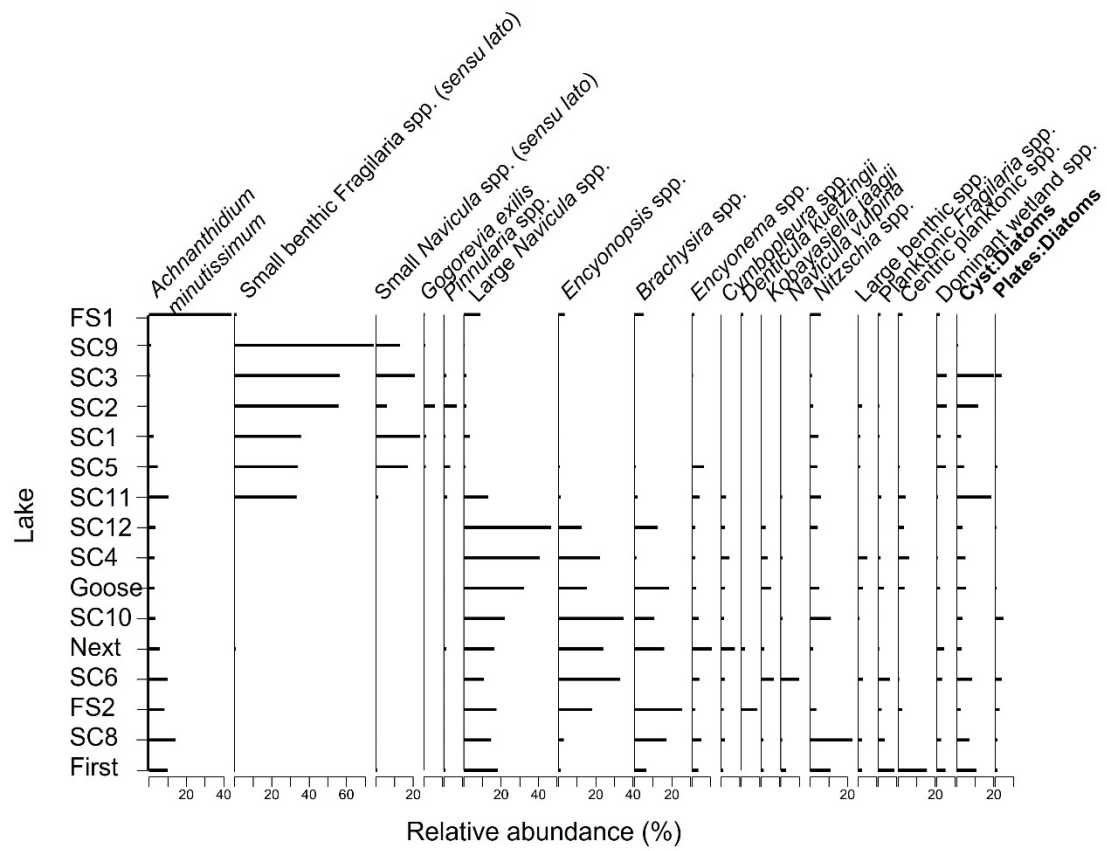
839

840

841

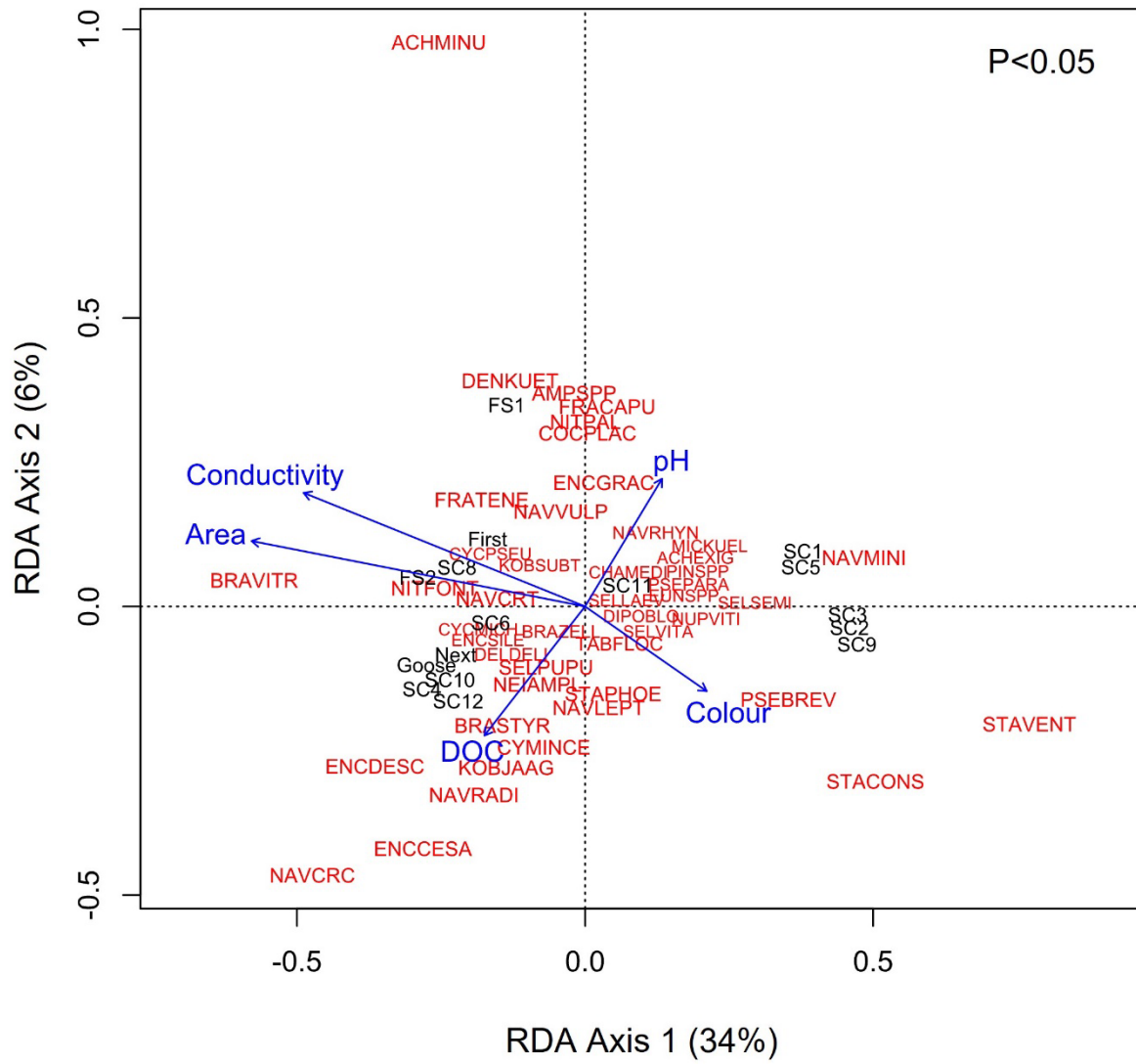
842

843

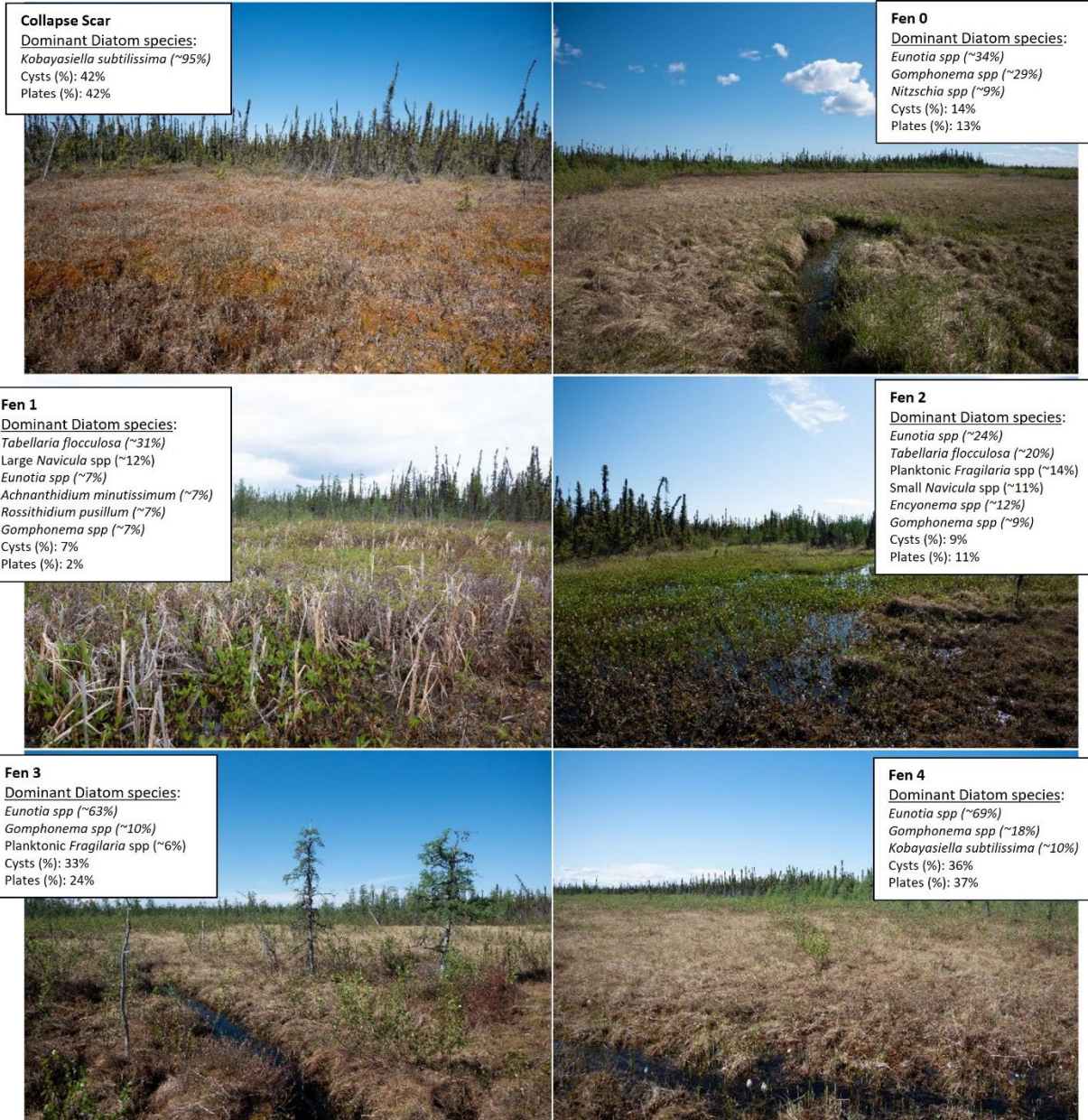


844

845



846



847

848 Appendices

849 Appendix A. List of diatom species groupings included in relative frequency diagram and taxon

850 codes used in RDA. Species present in low abundances (<2%) were not included in the RDA.

851 Some genera were grouped for RDA analysis (*Pinnularia* spp., *Eunotia* spp., *Amphora* spp.).

Groupings in relative frequency diagram	Species	Taxon Code
Small benthic Fragilariaceae	<i>Pseudostaurosira brevistriata</i>	PSEBREV



	<i>Pseudostaurosira parasitica</i>	PSEPARA
	<i>Staurosira construens</i>	STACONS
	<i>Staurosira construens var venter</i>	STAVENT
<b>Small <i>Navicula</i> spp. (sensu lato)</b>	<i>Sellaphora nigri</i>	NAVMINI
	<i>Sellaphora seminulum</i>	SELSEMI
<b><i>Encyonopsis</i> spp.</b>	<i>Encyonopsis cesatii</i>	ENCCESA
	<i>Encyonopsis descripta</i>	ENCDESC
<b><i>Pinnularia</i> spp.</b>	Grouped for RDA	PINSPP
	<i>Pinnularia gibba</i>	<2%
	<i>Pinnularia interrupta</i>	<2%
	<i>Pinnularia legumen</i>	<2%
	<i>Pinnularia maior/viridis</i>	<2%
	<i>Pinnularia microstauron</i>	<2%
	<i>Pinnularia nodosa</i>	<2%
	<i>Pinnularia obscura</i>	<2%
	<i>Pinnularia streptoraphe</i>	<2%
<b>Large <i>Navicula</i> spp.</b>	<i>Navicula cryptocephala</i>	NAVCRRC
	<i>Navicula cryptotenella</i>	NAVCRRT
	<i>Navicula radiosa</i>	NAVRADI
<b><i>Brachysira</i> spp.</b>	<i>Brachysira styriaca</i>	BRASTYR
	<i>Brachysira vitrea</i>	BRAVITR
	<i>Brachysira zellensis</i>	BRAZELL
<b><i>Encyonema</i> spp.</b>	<i>Encyonema gracilis</i>	ENCGRAC
	<i>Encyonema silesiacum</i>	ENCFILE
	<i>Encyonema hebridicum</i>	<2%
	<i>Encyonema perpusillum</i>	<2%
<b><i>Cymbopleura</i> spp.</b>	<i>Cymbopleura incerta</i>	CYMINCE
	<i>Cymbopleura angustata</i>	<2%
	<i>Cymbopleura lapponica</i>	<2%
<b><i>Nitzschia</i> spp.</b>	<i>Nitzschia fonticola</i>	NITFONT
	<i>Nitzschia palea</i>	NITPAL
	<i>Nitzschia alpina</i>	<2%
	<i>Nitzschia angustata</i>	<2%
	<i>Nitzschia intermedia</i>	<2%
	<i>Nitzschia recta</i>	<2%
	<i>Nitzschia vermicularis</i>	<2%
<b>Large benthic spp.</b>	<i>Neidium ampliatum</i>	NEIAMPL
	<i>Stauroneis phoenicenteron</i>	STAPHOE
	<i>Stauroneis javanica</i>	<2%
<b>Planktonic <i>Fragilaria</i> spp.</b>	<i>Fragilaria tenera</i>	FRATENE
	<i>Fragilaria nanana</i>	<2%
	<i>Fragilaria ulna</i>	<2%
<b>Centric planktonic spp.</b>	<i>Discostella pseudostelligera</i>	CYCPSEU
	<i>Lindavia michiganiana</i>	CYCMICH

	<i>Lindavia bodanica</i>	<2%
<b>Dominant wetland spp.</b>	<i>Gomphonema acuminatum</i>	<2%
	<i>Gomphonema angustatum</i>	<2%
	<i>Gomphonema gracile</i>	<2%
	<i>Gomphonema parvalum</i>	<2%
	<i>Gomphonema subtile</i>	<2%
	<i>Gomphonema truncatum</i>	<2%
	<i>Kobayasiella subtilissima</i>	KOBSUBT
	<i>Tabellaria</i> spp	TABFLOC
	<i>Eunotia</i> spp. (grouped for RDA)	EUNSPPP
	<i>Eunotia bilunaris</i>	<2%
	<i>Eunotia circumborealis</i>	<2%
	<i>Eunotia formica</i>	<2%
	<i>Eunotia incisa</i>	<2%
	<i>Eunotia praerupta</i>	<2%
	<i>Gogorevia exilis</i>	ACHEXIG
	<i>Achnantheidium minutissimum</i>	ACHMINU
	<i>Amphora</i> spp. (grouped for RDA)	AMPSPPP
	<i>Amphora pediculus</i>	<2%
	<i>Amphora thumensis</i>	<2%
	<i>Chamaepinnularia mediocris</i>	CHAMEDI
	<i>Cocconeis placentula</i>	COCPLAC
	<i>Delicata delicatula</i>	DELDELI
	<i>Denticula kuetzingii</i>	DENKUET
	<i>Diploneis oblongella</i>	DIPOBLO
	<i>Fragilaria capucina</i>	FRACAPU
	<i>Kobayasiella jaagii</i>	KOJAAG
	<i>Microcostatus kuelbsii</i>	MICKUEL
	<i>Navicula leptostriata</i>	NAVLEPT
	<i>Navicula rhychocephala</i>	NAVRHYN
	<i>Navicula vulpina</i>	NAVVULP
	<i>Nupela vitiosa</i>	NUPVITI
	<i>Sellaphora laevissima</i>	SELLAEV
	<i>Sellaphora pupula</i>	SELPUPU
	<i>Sellaphora vitabunda</i>	SELVITA